RESEARCH ARTICLE

Determination of Phenolics, Sugars, Organic Acids and Antioxidants in the Grape Variety Kalecik Karası under Different Bud Loads and Irrigation Amounts

Serpil Gök Tangolar[1*], Semih Tangolar[1], Haşim Kelebek[2], and Sevilay Topçu[3]

[1]Çukurova University, Faculty of Agriculture, Department of Horticulture, Adana, Turkey
[2]Adana Science and Technology University, Faculty of Engineering and Natural Sciences, Department of Food Technology, Adana, Turkey
[3]Cukurova University, Faculty of Agriculture, Department of Agricultural Structures and Irrigation, Adana, Turkey

*Corresponding author: stangolar@cu.edu.tr

Abstract

Irrigation applications (IA) and increased bud load (BL) are fundamental practices that are used to achieve optimum yields in grape production, while maintaining fruit quality parameters. Two different irrigation amounts (IA-I and IA-II) based on growth stages, in addition to a non-irrigated (rain-fed) control, along with two different BL applications [normal bud load based on traditional pruning practices (1BL) and double bud load (2BL)] were evaluated over a two-year experiment for their effects on the biochemical composition of the berries. Berries from the rain-fed vines had higher sugar levels, whereas no significant change was detected in organic acid levels. The increased bud load (2BL) treatment yielded less sugar in the berries compared to the 1BL control in both years. The total non-colored phenolic compounds (NPC) were greater in the irrigated (especially IA-I) and 2BL treatments than in those of the rain-fed and 1BL control. However, total anthocyanin was greater in the non-irrigated and 1BL control than in the irrigated and 2BL treatments. The antioxidant contents of the berries also varied according to the treatments and years. Our results implied that implementing a higher bud load along with the IA-I irrigation application, in which irrigation applications were 50 and 75% of the cumulative evaporation from the Class A pan during berry set to veraison and veraison to harvest growth stages, respectively, can help in obtaining greater yields in high-plateau viticulture. Thus, if more buds are left on the vines, along with sufficient irrigation and rainfall, yield may increase while maintaining or increasing the biochemical composition of the berries.

Additional key words: biochemical composition, catechin, epicatechin, grapevine, malvidin

Introduction

Grape production is increasing world-wide due to the benefits that grape consumption has for human health and nutrition. The biochemical composition of the grapes is the main determinant of

quality. Sugars, organic acids, and phenolic compounds are the main constituents in grapes (Zhu et al., 2012; Eshghi et al., 2014; Rahmani et al., 2015). Grapes have a high adaptability to different conditions and are used for various purposes such as direct consumption of the fruits and for making wine, raisins, vinegar, molasses, and juice, all of which add economic value to grape production.

Although the nutritional value of grapes is high, consumers demand not only aesthetically pleasing fruit, but also environmentally friendly production. Therefore, recent research has focused on the analysis of the biochemical compounds (Beslic et al., 2010; Wessner and Kurtural, 2013; Kurtural et al., 2013) and the antioxidant contents of grapes (Zhu et al., 2012; Rahmani et al., 2015). Many studies have documented large amounts of antioxidants and phenolic compounds in table grapes, wines, raisins, and other grape products (Breksa et al., 2010; Göktürk Baydar et al., 2011; Kelebek et al., 2013; Wessner and Kurtural, 2013). Recent research has reported that phenolic compounds have antioxidant, anti-allergenic, anti-cancer, anti-microbial, anti-ageing, and anti-inflammatory properties (Beslic et al., 2010; Rahmani et al., 2015). The phenolic compounds in grapes are divided into colored phenols, such as anthocyanins and flavonoids, and non-colored phenols, including phenolic acids, stilbenes, and flavan-3-ols (Silva et. al., 2005; Sun et al., 2007; Kelebek, 2009; Kelebek et al. 2013). The content of phenolic compounds in grapes is affected by variety, maturity, and several environmental and cultural factors including the vine-training system, biotic and abiotic stress conditions, crop load, irrigation, fertility, and summer pruning, among others (Kennedy et al., 2002; Beslic et al., 2010; Fawzi et al., 2010; Terry and Kurtural, 2011; Kurtural et al., 2013).

Pruning is an essential cultural practice in grape production. It affects the form and size of the vine and establishes a balance between shoot and fruit growth. Hence, the level of pruning has an effect on yield, quality, and fruit composition (Kurtural et al., 2013; Wessner and Kurtural, 2013; Rahmani et al., 2015) due to altered canopy size, shoot development, and fruit cluster numbers per vine.

Irrigation also affects the quality of the grapes. In locations with an annual rainfall of 500-600 mm, rain-fed viticulture can be successful depending on the soil type. However, supplementary irrigation might be necessary to maintain high yield and quality when the seasonal distribution of rainfall is not adequate (Çelik, 2011).

Kalecik Karası (*Vitis vinifera* L.) is one of the most important Turkish grape cultivars grown for red wine production. It originated from Kalecik, a town in the Ankara province, mid-north Turkey. Information on the effects of irrigation and pruning to manage fruit load is lacking for this variety. The objective of this study was to evaluate the effect of different irrigation applications (IA) and bud loads (BL) on the biochemical composition of Kalecik Karası grown in the high plateau regions.

Materials and Methods

Berries from own rooted sixteen-year-old grape vines of cv. Kalecik Karası (*Vitis vinifera* L.) grown in the Pozantı Agricultural Research Centre (1,080 m above sea level) (37°28'38.1"N, 34°54'09.5"E) of Cukurova University (Adana, Turkey) were used as the plant material in 2013 and 2014. The vines, grown in 2m × 3m (vine x row) plots in an east-west orientation, were trained to a bilateral cordon system. The soil in the experimental vineyard was slightly alkaline (pH 7.64-7.77) with clay-loam and loam textures on the surface and at a 30-60 cm soil depth. The field capacity and the permanent wilting point (average values for the two soil depths) were 25.7 and 17.5%, respectively. Climatic data were obtained from

the Climate Station of Regional Directorate of Meteorology in Pozantı, Turkey. In April through October of 2013 and 2014, the average daily temperatures were 19.1 and 18.5℃, relative humidity was 45.8 and 59.2% and total precipitation was 135.0 and 321.2 mm with a 2.2 and 4.4 m·s⁻¹ wind speed, respectively.

Three treatments were used in the experiment, one increased bud load treatment and a normal bud load control and two irrigation treatments and a non-irrigated (rain-fed) control.

Bud Load Levels

Vines maintained at the normal bud load (1BL, control) according to traditional practices (20 buds for the first 500 g of pruning weights and 10 more buds for every subsequent 500 g of pruning weight) (Winkler et al., 1974; Çelik, 2011) and increased bud loads (2BL, double the buds of the control) (Table 1) were used for all irrigation treatments. After dormant pruning, the average pruning weights to determine the number of buds to be retained were measured on March 8th and 10th in 2013 and 2014, respectively.

Table 1. Retained bud number by pruning weight in irrigation amounts (IA)[z] and bud load (BL) treatments of Kalecik Karası

	Pruning weight (g/vine)	Retained bud number (n) for 1BL
	2013	
For all treatments	500[y]	20
	2014	
Source of variance[x]		
Water amounts		
No irrigation	682 b[w]	23
IA-I	938 a	28
IA-II	1068 a	31
(Pr > F)	** [v]	
Bud load		
1BL	876	27
2BL	917	28
(Pr > F)	NS	
IWA × BL		
No irr. × 1BL	627	22
No irr. × 2BL	737	24
IA-I. × 1BL	957	29
IA-I × 2BL	920	28
IA-II. × 1BL	1043	30
IA-II × 2BL	1093	32
(Pr > F)	NS	

[z]Values for the water amount main effects are the mean of 4 measurements (2 bud loads and 2 replicates) and values for the bud load main effects are the mean of 6 measurements (3 water amounts and 2 replicates).

[y]This value was the same for all treatments.

[x]1BL (control) represents retaining 20 buds for the first 500 g of pruning weight plus 10 more buds for each additional 500 g., 2BL: retained double the buds of 1BL, irr.: Irrigation.

[w]Means followed by different letters on the same column are significantly different according to LSD test at $p \leq 0.05$.

[v]NS, *, ** or ***: Nonsignificant or significant at $p \leq 0.05$, 0.01, or 0.001, respectively.

Irrigation Treatments

In both of the irrigation treatments, the timing of irrigation was scheduled according to the midday leaf water potential (LWP), a widely used measurement of plant water status, while the irrigation volume applied was calculated using the cumulative evaporation (Epan) values obtained from Class-A Evaporation Pans (METOS Inc., Mersin, Turkey). The midday LWP values that dictated when irrigation was to be applied differed for each plant growth stage: -10 bars for the period before flowering, -13 bars for the period between fruit set and ripening and -12 bars for the post-ripening stage. Water amounts applied to the IA-I plots were 50% of the cumulative evaporation from the Class A Pan for the berry setting through veraison growth stage and 75% for the veraison through harvest growth stage. For the IA-II plots, the water amounts applied were 75% of the evaporation from the Pan for the berry setting through veraison growth stage and 50% for the veraison through harvest growth stage. The irrigation water amount was calculated using the following equation based on cumulative Class-A pan evaporation within the irrigation intervals.

$$I = A \times Epan \times Kpc \times P$$

I: Irrigation volume (mm or L), A: parcel area (6 m^2), Epan: Evaporation from the class-A pan (mm or L), Kpc: The crop and pan coefficient (0.6) and P: Percentage of wetted area (0.5). P value was calculated as the percentage of the average area shaded by the canopy measured at noon. Drip lines with an emitter spaced every 20 cm were placed close to the vine rows and were operated at a 2 L/h flow rate and a pressure of 1.5 bars. Irrigation amounts and dates of irrigation are presented in Fig. 1.
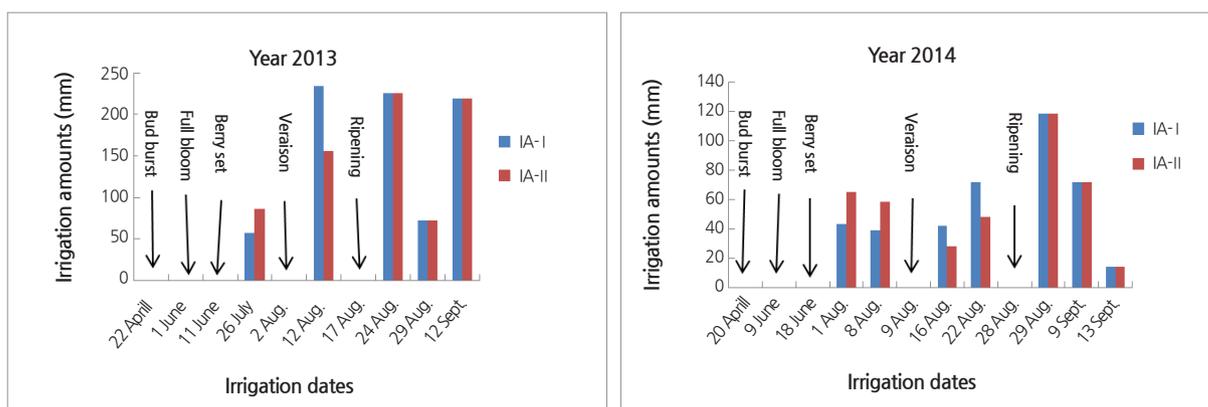
Fig. 1. Irrigation dates and amounts (mm) in the different irrigation treatments

Four fully expanded leaves exposed to direct sunlight were chosen for the leaf water potential (LWP) measurement in each plot. To measure midday LWP (between 11:30 and 14:00 hrs), the targeted leaves were first covered entirely with a small plastic bag and then the petiole of the bagged leaf was cut from the shoot with a sharp blade. A pressure chamber device (Model 600, PMS Instrument Co. Albany, Oregon, USA) was used for the LWP measurements. Stomatal conductance and leaf temperature were measured using a leaf porometer (Model SC-1, Decagon devices Inc., Pullman, WA 99163, USA) on two fully expanded leaves exposed to direct sunlight and light intensity was measured using a Light Meter (Model LX-1102, Lutron Electronic Enterprise Co. Ltd., Taipei, TAIWAN). During harvest (when the amount of total soluble solids (TSS) has reached approximately 22-23%) 500 g of berries with pedicels were taken from the inner two vines with two

replicates. These samples were stored at -80°C until the following analyses were performed.

Analysis of Sugars and Organic Acids

To determine sugars and organic acids, samples were prepared according to the method proposed by Sturm et al. (2003). Briefly, 100 g of whole fruit without pedicel was converted into pulp by a mixer and homogenized with an Ultra-Turrax T-25 (IKA Laboratory Equipment). The fruit puree (10 g) was diluted to 100 mL with double-distilled water and centrifuged at 5,000 rpm for 15 min. The extract was filtered through 0.45 μm Millipore filters and a 20 μL sample was used for HPLC analysis of sugars and organic acids. An Agilent 1100 HPLC system (Agilent Technologies, Palo Alto, CA, USA) equipped with a pump system, a refractive index detector for sugar analysis, and a diode array detector was monitored at 210 nm for the analysis of organic acids. Sugars and organic acids were simultaneously analysed using an Aminex HPX-87H column (300 × 7.8 mm) (Bio-Rad, UK) and maintained at 55°C. The analytical conditions used were as follows, flow: 0.3 mL·min^{-1}, eluent: 0.045 NH_2SO_4 with 6% acetonitrile (v/v). The chromatographic peaks corresponding to each sugar and organic acid were identified by comparing the retention times with that of a standard.

Analysis of Phenolic Compounds

Extracts for analysis of phenolic compounds were prepared from whole fruits without pedicels according to the method of Breksa et al. (2010). An Agilent 1100 HPLC system, as described above, with a diode array detector operated by Windows NT-based ChemStation software was used. Separation was performed on a Beckman Ultrasphere ODS column (Roissy, France; 4.6 mm × 250 mm, 5 μm). The mobile phase consisted of water with 5% formic acid (solvent A; v/v) and acetonitrile with 40% solvent A (solvent B; v/v). The elution program was conducted as previously described (Kelebek et al., 2013). The identification and determination of each compound was done by comparing their retention times and UV spectra to standards and also confirmed by an Agilent 6430LC-MS/MS spectrometer equipped with an electrospray ionization source. The electrospray ionization mass spectrometry detection was performed in negative ion mode with the following optimized parameters: capillary temperature 400°C, capillary voltage -3V, nebulizer gas flow 1.75 L·min^{-1}, desolation gas flow 1 L·min^{-1} and spray voltage 5 kV. The analysis was conducted in negative and positive modes (Kelebek et al., 2013). Data gaining was achieved in the multiple reaction monitoring (MRM) mode that only monitors specific mass transitions during preset retention times. Dynamic MRM also has the ability to simultaneously carry through electrospray ionization in both positive modes (for anthocyanins) and negative modes (for phenolic acids, flavonols and flavanols), allowing for the analysis of different phenolic classes within a single chromatographic run.

Analysis of Antioxidants

The electron donation ability of the fruit extract was measured by bleaching the purple-colored solution of 1,1-diphenyl-2-picrylhydrazyl (DPPH) radical described by Kelebek et al. (2013). Three different dilutions of the grape extracts (1, 20 and 50 mg·mL^{-1}) were prepared in ethanol/water (4/1, v/v). An aliquot of 0.1 mL of diluted grape extract was added to 3.9 mL of DPPH solution in methanol (6×10^{-5} M). The mixture was shaken vigorously and left at room temperature for 30 min. The absorbance of the resulting solution was then measured at 515 nm by a Cary 60 UV-Vis spectrophotometer (Agilent Technologies, Palo Alto, CA, USA) until the reaction reached the plateau (steady state).

Statistical Analysis

The experiment was designed as a split plot with two replicates. Each experimental unit consisted of five vines. Twelve samples were used for each attribute analysed. The irrigation treatments were the main plots and the bud load treatments were sub plots. Analysis of variance (ANOVA) was done using the JMP computer program for statistics developed by the JMP business unit of SAS Institute and the least significant difference (LSD) test was used for separation of the means at a 5% significance level. Interactions between year and treatments were tested and analysis was conducted separately whenever these interactions were significant ($p \leq 0.05$).

Results and Discussion

The applied irrigation volume (IA) in different periods for each treatment is given in Figure 1. The first irrigation in 2013 was applied on July 26 and in 2014 the first irrigation was applied on August 1. LWP values were below the threshold of -10 bars until berry set and -13 bars after berry set; therefore, the recorded 247 mm and 437.4 mm of precipitation from January to August in 2013 and 2014, respectively, (Fig. 2) was considered sufficient for grape cultivation (Winkler et al., 1974; Weaver, 1976; Ward, 1984; Çelik et al., 1998; Çelik, 2011). Ward (1984) suggested that for successful vine production, average annual rainfall should not exceed 762 mm, and during the growing period precipitation should remain below 350 mm. Winkler et al., (1974) indicated that some varieties of grapes can be grown in cool regions with 381 mm of water, and occasionally less. Similarly, Weaver (1976) stated that annual rainfall varies between 300-800 mm in cooler regions of California and Arizona. Çelik et al. (1998) and Çelik (2011) also stated that grape vines can be successfully grown in areas with an average annual precipitation between 450-500 mm.

Glucose, fructose (except in 2013), and total sugars (Table 2) were highest in the non-irrigated control. The lowest values for glucose, fructose, and total sugars were found in the IA-II treatment (except in 2014) where more water was applied
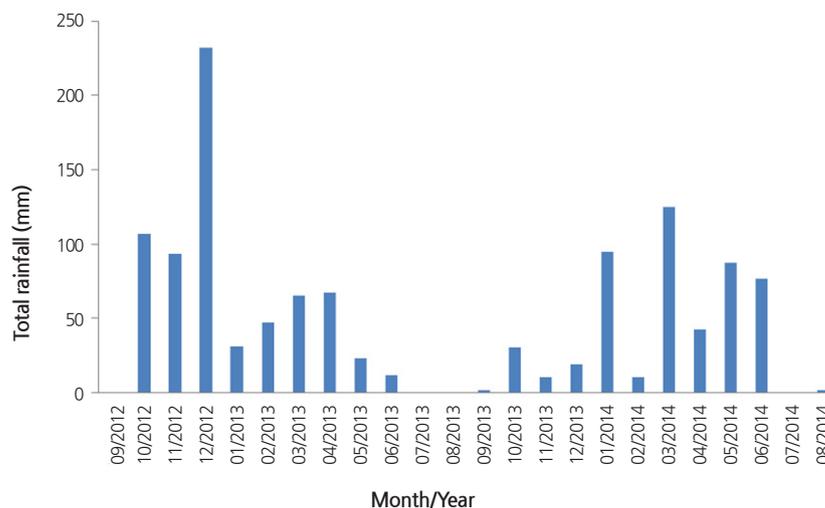
Fig. 2. Total monthly rainfall between September, 2012 and August, 2014 in the experimental plots (Regional Directorate of Me -teorology, Climate Station, Pozantı, south Turkey)

between berry set and veraison (85.7 mm/vine and 123.8 mm/vine for 2013 and 2014, respectively) than in the IA-I treatment. Glucose, fructose, and total sugars were also significantly higher ($p \leq 0.05$) in the traditional BL control compared to those in the 2BL (increased BL) treatment in both years. Our results were in accordance with other studies that reported that excessive bud load can reduce available sugars and water absorption in the berries (Winkler et al., 1974; Cus et al., 2004; Çelik, 2011). Total soluble solids were also not affected by pruning treatments in Wilson et al. (2014). Sucrose was very low compared to other sugar components. Both IA and BL treatments had no effect on tartaric, malic and total organic acids of 'Kalecik Karası' berries in either year of the our study. Cus et al. (2004) also found a similar effect of the crop load in their research.

In total, thirteen anthocyanin compounds were detected in the grapes from our study. Five were monoglucosides, four were acetylated and the remaining four compounds were coumarylated (Table 3). Considering all of the anthocyanin

Table 2. Effects of irrigation amount (IA)[z] and bud load (BL) on sugar and organic acid contents in Kalecik Karası

Source of variance[y]	Glucose	Fructose	Sucrose	Total sugars	Tartaric acid	Malic acid	Total acids
	(g·kg⁻¹ fresh weight)						
	2013						
Water amounts							
No irrigation	104.87 a[x]	102.97 a	1.11 a	208.96 a	4.97	2.53	7.50
IA-I	98.27 b	99.27 a	0.27 b	197.81 b	5.40	2.97	8.36
IA-II	93.05 c	92.79 b	1.11 a	186.95 c	4.85	3.43	8.28
(Pr > F)	**[w]	**	***	**	NS	NS	NS
Bud load							
1BL	100.97 a	100.96 a	0.00 b	201.94 a	5.18	2.80	7.98
2BL	96.49 b	95.72 b	1.66 a	193.87 b	4.96	3.15	8.11
(Pr > F)	*	*	***	*	NS	NS	NS
Irr. × BL (Pr > F)	*	*	***	*	NS	NS	NS
	2014						
Water amounts							
No irrigation	104.56 a	94.93 a	2.81 a	202.30 a	4.89	3.03	7.93
IA-I	97.30 b	88.85 b	2.67 a	188.81 b	4.76	2.93	7.69
IA-II	94.99 b	87.91 b	2.46 b	185.36 b	5.09	2.77	7.86
(Pr > F)	*	*	*	*	NS	NS	NS
Bud load							
1BL	102.42 a	93.42 a	2.78 a	198.63 a	4.81	3.02 a	7.84
2BL	95.48 b	87.71 b	2.51 b	185.69 b	5.02	2.80 b	7.82
(Pr > F)	**	*	*	**	NS	*	NS
Irr. × BL (Pr > F)	*	*	NS	*	NS	NS	NS
Year (Pr > F)	NS	***	***	**	NS	NS	NS
Year × Irr. (Pr > F)	NS	*	***	NS	NS	NS	NS
Year × BL (Pr > F)	NS	NS	***	NS	NS	NS	NS
Year × Irr. × BL (Pr > F)	**	***	**	***	NS	NS	NS

[z]Values for the water amount main effects are the mean of 4 measurements (2 bud loads and 2 replicates) and values for the bud load main effects are the mean of 6 measurements (3 water amounts and 2 replicates).

[y]1BL (control) represents retaining 20 buds for the first 500 g of pruning weight plus 10 more buds for each additional 500 g., 2BL: retained double the buds of 1BL, irr.: Irrigation.

[x]Means followed by different letters on the same column are significantly different according to LSD test at $p \leq 0.05$.

[w]NS, *, ** or ***: Nonsignificant or significant at $p \leq 0.05$, 0.01, or 0.001, respectively

compounds (except delphinidin 3-glucoside), the significantly highest values were obtained from the grapevines grown under the rain-fed conditions ($p \leq 0.05$) followed by the IA-II treatment. Total anthocyanin content was also greater in the non-irrigated berries in previous reports (Esteban et al., 2001; Chaves et al., 2007; Özden et al., 2010; Santesteban et al., 2011), but not in Zarrouk et al. (2012). According to the results of Kyraleou et al. (2015) water restriction had positive effects on total anthocyanin levels, which has also been noted in their varieties (Castellarin et al., 2007; Bucchetti et al., 2011). The normal BL level based on traditional practices resulted in greater amounts of total anthocyanin in both years of the study (Table 3). Increases in the anthocyanin concentration in the berries from the rain-fed and the 1BL controls may be affected by changes in leaf stomatal conductance, leaf temperature, and light intensity within the canopy (Table 4) (Downey et al., 2004) or from direct effects on the expression of genes of the flavonoid pathway (Castellarin et al., 2007). Kurtural et al. (2013) also showed that anthocyanins in berry skin decreased with a 45 shoots/m treatments when compared with 40

Table 3. Effects of irrigation amount (IA)[z] and bud load (BL) on anthocyanin contents in Kalecik Karası

Source of variance[x]	Del 3glu[y]	Cyanidin 3glu	Pet 3glu	Peonidin 3glu	Malvidin 3glu	Del3 gluacet	Pet3 gluacet
	(mg·kg^{-1} fresh weight)						
	2013						
Water amounts							
No irrigation	2.77 b[w]	1.06 a	6.41 a	12.58 a	59.80 a	1.50 a	2.24 a
IA-I	3.00 a	0.64 c	4.19 c	6.89 c	46.53 c	1.17 c	1.93 c
IA-II	1.90 c	0.84 b	5.05 b	7.77 b	51.68 b	1.30 b	2.03 b
(Pr > F)	***[v]	***	***	***	***	***	***
Bud load							
1BL	2.69 a	0.87 a	4.89 b	9.14 a	51.38 b	1.26 b	2.01 b
2BL	2.41 b	0.82 b	5.54 a	9.02 b	53.96 a	1.39 a	2.13 a
(Pr > F)	***	***	***	*	***	***	***
Irr. × BL (Pr > F)	***	***	***	***	***	***	***
	2014						
Water amounts							
No irrigation	1.74 b	0.29 a	4.70 a	10.40 a	50.57 a	0.52 c	2.42 a
IA-I	1.86 a	0.05 c	2.92 c	4.94 c	36.82 c	0.89 b	1.74 b
IA-II	0.99 c	0.21 b	3.54 b	5.64 b	42.65 b	0.97 a	1.42 c
(Pr > F)	***	**	***	***	***	***	***
Bud load							
1BL	1.66 a	0.19	3.47 b	7.25	42.82	0.72 b	1.93 a
2BL	1.40 b	0.18	3.97 a	6.75	43.88	0.87 a	1.79 b
(Pr > F)	***	NS	***	NS	NS	***	***
Irr. × BL (Pr > F)	***	*	***	**	**	***	***
Year (Pr > F)	***	***	***	***	***	***	***
Year × Irr. (Pr > F)	NS	***	NS	NS	NS	NS	NS
Year × BL (Pr > F)	NS	NS	NS	NS	NS	NS	NS
Year × Irr. × BL (Pr > F)	NS	***	NS	NS	NS	NS	NS

[z]Values for the water amount main effects are the mean of 4 measurements (2 bud loads and 2 replicates) and values for the bud load main effects are the mean of 6 measurements (3 water amounts and 2 replicates).

[y]3glu: 3 glucoside, Del: Delphinidin, Pet: Petunidin, acet: acetyl

[x]1BL (control) represents retaining 20 buds for the first 500 g of pruning weight plus 10 more buds for each additional 500 g., 2BL: retained double the buds of 1BL, irr.: Irrigation.

[w]Means followed by different letters on the same column are significantly different according to LSD test at $p \leq 0.05$.

[v]NS, *, ** or ***: Nonsignificant or significant at $p \leq 0.05$, 0.01, or 0.001, respectively.

Table 3(Continued). Effects of irrigation amount (IA)[z] and bud load (BL) on antochyanin contents in Kalecik Karası

Source of variance[x]	Peo 3gluacet[y]	Malv 3gluacet	Del3 glucoum	Pet3 glucoum	Peo 3 glucoum	Malv3 glucoum	Total
	(mg·kg^{-1} fresh weight)						
	2013						
Water amounts							
No irrigation	1.61 a[w]	19.99 a	0.83 a	0.87 a	2.37 a	19.90 a	131.95 a
IA-I	0.48 c	17.36 b	0.80 b	0.41 c	1.10 c	16.68 b	101.18 c
IA-II	0.66 b	16.85 c	0.73 c	0.56 b	1.21 b	16.34 c	106.91 b
(Pr > F)	***[v]	***	***	***	***	***	***
Bud load							
1BL	0.97 a	18.68 a	0.83 a	0.68 a	1.74 a	18.70 a	113.82 a
2BL	0.87 b	17.46 b	0.75 b	0.55 b	1.38 b	16.58 b	112.87 b
(Pr > F)	***	***	***	*	***	***	***
Irr. × BL (Pr > F)	***	***	***	***	***	***	***
	2014						
Water amounts							
No irrigation	1.31 a	17.00 a	0.67 a	0.72 a	1.94 a	15.71 a	108.08 a
IA-I	0.39 c	13.48 b	0.63 b	0.33 c	0.87 c	12.95 c	78.56 c
IA-II	0.52 b	13.03 b	0.58 c	0.44 b	0.95 b	13.68 b	84.92 b
(Pr > F)	***	**	***	***	***	**	***
Bud load							
1BL	0.79 a	15.20 a	0.67 a	0.55 a	1.42 a	15.10 a	91.78 a
2BL	0.70 b	13.81 b	0.59 b	0.44 b	1.09 b	13.12 b	89.26 b
(Pr > F)	**	*	***	**	***	**	*
Irr. × BL (Pr > F)	***	*	***	***	***	**	**
Year (Pr > F)	***	***	***	***	***	***	***
Year × Irr. (Pr > F)	**	NS	NS	NS	NS	NS	NS
Year × BL (Pr > F)	NS	NS	NS	NS	NS	NS	NS
Year × Irr. × BL (Pr > F)	NS	NS	NS	NS	NS	NS	NS

[z]Values for the water amount main effects are the mean of 4 measurements (2 bud loads and 2 replicates) and values for the bud load main effects are the mean of 6 measurements (3 water amounts and 2 replicates).

[y]Del: Delphinidin, Pet: Petunidin, Peo: Peonidin, Malv: Malvidin, 3glu: 3 glucoside, acet: acetyl, coum: coumaryl

[x]1BL (control) represents retaining 20 buds for the first 500 g of pruning weight plus 10 more buds for each additional 500 g., 2BL: retained double the buds of 1BL, irr.: Irrigation.

[w]Means followed by different letters on the same column are significantly different according to LSD test at $p \leq 0.05$.

[v]NS, *, ** or ***: Nonsignificant or significant at $p \leq 0.05$, 0.01, or 0.001, respectively.

shoots/m treatments.

Water stress or limited water led to a less-dense canopy, which may have resulted in greater irradiance within the fruit zone of the rain-fed plots. The non-irrigated plants also had greater leaf temperature and lower stomatal conductance compared to the irrigated plants. Accordingly, higher sugar and anthocyanin contents have been found in grape berries of plants under rain-fed conditions. Previous studies (Shellie, 2011; Terry and Kurtural, 2011; Zarrouk, et al., 2012; Nelson et al., 2015) showed that water deficits affected the physiological and hormonal responses of field and glasshouse grown grapevines. Generally, the less water applied, the more stomatal conductance was reduced. Bindon et al. (2011) reported that frequent irrigation can cause a remarkable restriction of anthocyanin biosynthesis. Chalmers (2007) examined the effects

Table 4. Effects of irrigation amount (IA)[z] and bud load (BL) on leaf stomatal conductance, leaf temperature and canopy light intensity

Source of variance[y]	Stomatal conductivity (mmol·m^{-2}·s^{-1})	Leaf temperature (°C)	Light intensity within canopy (lux)	Light intensity outside canopy (lux)
		2013		
Water amounts				
No irrigation	603.0 b[x]	35.4 a	3185 a	111357±2710 [v]
IA-I	942.1 a	34.5 b	2532 b	
IA-II	1023.5 a	34.4 b	2371 b	
(Pr > F)	**[w]	*	*	
Bud load				
1BL	817.4	34.9 a	2601	
2BL	895.0	34.6 b	2790	
(Pr > F)	NS	*	NS	
Irr. × BL (Pr > F)	NS	*	NS	
		2014		
Water amounts				
No irrigation	380.8 c	37.1 a	2396 a	113983±1605
IA-I	749.9 a	37.0 a	1899 b	
IA-II	656.0 b	36.7 b	1910 b	
(Pr > F)	***	*	**	
Bud load				
1BL	600.3	37.0	2045	
2BL	590.9	36.9	2092	
(Pr > F)	NS	NS	NS	
Irr. × BL (Pr > F)	***	NS	*	
Year (Pr > F)	***	***	**	
Year × Irr. (Pr > F)	*	***	NS	
Year × BL (Pr > F)	NS	*	NS	
Year × Irr. × BL (Pr > F)	NS	*	NS	

[z]Values for the water amount main effects are the mean of 4 measurements (2 bud loads and 2 replicates) and values for the bud load main effects are the mean of 6 measurements (3 water amounts and 2 replicates). For each replicate, the mean of 8 measurements performed on different dates in July and August was taken.
[y]1BL (control) represents retaining 20 buds for the first 500 g of pruning weight plus 10 more buds for each additional 500 g., 2BL: retained double the buds of 1BL, irr.: Irrigation.
[x]Means followed by different letters on the same column are significantly different according to LSD test at $p \leq 0.05$.
[w]NS, *, ** or ***: Nonsignificant or significant at $p \leq 0.05$, 0.01, or 0.001, respectively.
[v]These values were provided for information purposes to the readers as the mean of the 8 measurements done in July and August.

of different levels of irrigation on grapes and also found total anthocyanin between 1.77 and 1.07 mg·g^{-1} for full (100%) irrigation but found higher values (1.22 and 2.25 mg·g^{-1}) in limited water (45% of full irrigation) conditions. In our study, total anthocyanin varied between 101.18 mg·kg^{-1} (IA-I) and 131.95 mg·kg^{-1} (rain-fed) for 2013 and 78.56 mg·kg^{-1} (IA-I) and 108.08 mg·kg^{-1} (rain-fed) for 2014 (Table 3), which were close to the values found by Cantos et al. (2002). Anthocyanins were the main phenolic compounds in red grapes ranging from 151 to 689 mg·kg^{-1} fresh berry weight. Previous studies reported higher anthocyanin values, such as between 471 and 1,242 mg kg^{-1} (Bindon et al., 2011) and 940 to 1,270 mg·kg^{-1} (Chalmers, 2007). The anthocyanin values in the skin of the berries varied between 355.9 and 902.1 mg·kg^{-1} for the Syrah variety in Wessner and Kurtural (2013). Eshghi et al. (2014) also reported that the total anthocyanin content of berries varied significantly among the 35 grape cultivars grown in Iran. They indicated that the total content of anthocyanin in the berries

Table 5. Effects of irrigation amount (IA)[z] and bud load (BL) on flavanol contents in Kalecik Karası

Source of variance[y]	Catechin	Epicatechin
	(mg·kg⁻¹ fresh weight)	
	2013	
Water amounts		
No irrigation	130.68 c[x]	170.53 c
IA-I	145.39 a	255.45 a
IA-II	142.06 b	248.44 b
(Pr > F)	***[w]	***
Bud load		
1BL	130.17 b	203.44 b
2BL	148.59 a	246.17 a
(Pr > F)	***	***
Irr. × BL (Pr > F)	***	***
	2014	
Water amounts		
No irrigation	147.87 c	171.27 c
IA-I	157.83 a	256.55 a
IA-II	154.21 b	249.52 b
(Pr > F)	**	***
Bud load		
1BL	145.31 b	204.32 b
2BL	161.30 a	247.24 a
(Pr > F)	***	***
Irr. × BL (Pr > F)	***	***
Year (Pr > F)	***	***
Year × Irr. (Pr > F)	***	***
Year × BL (Pr > F)	***	***
Year × Irr. × BL (Pr > F)	***	***

[z]Values for the water amount main effects are the mean of 4 measurements (2 bud loads and 2 replicates) and values for the bud load main effects are the mean of 6 measurements (3 water amounts and 2 replicates).

[y]1BL (control) represents retaining 20 buds for the first 500 g of pruning weight plus 10 more buds for each additional 500 g., 2BL: retained double the buds of 1BL, irr.: Irrigation.

[x]Means followed by different letters on the same column are significantly different according to LSD test at $p \leq 0.05$.

[w]NS, *, ** or ***: Nonsignificant or significant at $p \leq 0.05$, 0.01, or 0.001, respectively.

of grapevine cultivars varied from 42.74 mg·kg⁻¹ to 619.04 mg·kg⁻¹ based on fresh weight. The order of amounts of non-acylated anthocyanins in both years was malvidin > peonidin > petunidin > delphinidin > cyanidin, consistent with previous results obtained for different native Greek varieties (Kallithraka et al., 2005) and for Syrah grapes grown under semiarid conditions (Kyraleou et al., 2015). The cyanidin level was as low as 1% of total anthocyanins (Winkler et al., 1974; Kelebek, 2009). Although our values are partially in agreement with those reported in previous studies, the differences may result from variety, maturation stage, ecological conditions, viticultural practices, and different analysis methods (Cantos et al., 2002; Chalmers, 2007; Kelebek 2009; Lachman et al., 2009; Kurtural et al., 2013).

For the non-colored phenolic compounds (NPC), two flavanols (Table 5), seven phenol acids (Table 6), and four flavonols (Table 7) were identified and quantified in Kalecik Karası berries. All of these compounds were affected by the treatments in both years of the study. The levels of flavanols, some phenolic acids (gallic and ferulic), and total NPC were increased by the irrigation treatments in both years. Catechin and epicatechin (Table 5) were significantly higher ($p \leq 0.05$) in the IA-I

Table 6. Effects of irrigation amount (IA)[z] and bud load (BL) on phenol acid contents in Kalecik Karası

Source of variance[y]	Gallic acid	Protoca techuic acid	Trans caftaric acid	Trans coutaric acid	Caffeic acid	p-coumaric acid	Ferulic acid
				(mg·kg⁻¹ fresh weight)			
				2013			
Water amounts							
No irrigation	11.49 c[x]	11.76 a	50.55 a	14.14 a	0.27 a	0.93 a	1.14 c[x]
IA-I	12.26 b	9.05 c	39.69 c	10.61 c	0.00 b	0.90 b	1.51 b
IA-II	13.51 a	9.58 b	47.01 b	13.36 b	0.00 b	0.83 c	1.85 a
(Pr > F)	***[w]	***	***	***	***	***	***[w]
Bud load							
1BL	12.01 b	9.41 b	40.73 b	12.17 b	0.18 a	0.81 b	1.42 b
2BL	12.82 a	10.85 a	50.77 a	13.24 a	0.00 b	0.96 a	1.59 a
(Pr > F)	***	***	***	***	***	***	***
Irr. × BL (Pr > F)	***	***	***	***	***	***	***
				2014			
Water amounts							
No irrigation	9.91 c	8.94 a	79.51 a	16.78 a	0.53 a	0.99 a	1.22 c
IA-I	10.57 b	6.89 c	62.44 c	12.60 c	0.25 b	0.96 b	1.61 b
IA-II	11.65 a	7.29 b	72.20 b	15.86 b	0.13 c	0.88 c	1.97 a
(Pr > F)	***	***	**	***	**	***	***
Bud load							
1BL	10.36 b	7.16 b	64.07 b	14.44 b	0.35 a	0.87 b	1.51 b
2BL	11.06 a	8.25 a	78.70 a	15.72 a	0.26 b	1.02 a	1.69 a
(Pr > F)	***	***	**	***	*	***	***
Irr. × BL (Pr > F)	***	***	**	***	NS	***	***
Year (Pr > F)	***	***	***	***	***	***	***
Year × Irr. (Pr > F)	***	***	***	***	***	***	***
Year × BL (Pr > F)	***	***	***	***	***	***	***
Year × Irr. × BL (Pr > F)	***	***	***	***	***	***	***

[z]Values for the water amount main effects are the mean of 4 measurements (2 bud loads and 2 replicates) and values for the bud load main effects are the mean of 6 measurements (3 water amounts and 2 replicates).

[y]1BL (control) represents retaining 20 buds for the first 500 g of pruning weight plus 10 more buds for each additional 500 g., 2BL: retained double the buds of 1BL, irr.: Irrigation.

[x]Means followed by different letters on the same column are significantly different according to LSD test at $p \leq 0.05$.

[w]NS, *, ** or ***: Nonsignificant or significant at $p \leq 0.05$, 0.01, or 0.001, respectively.

treatment than in the other treatment and the control for both years of the study. Gallic acid, ferulic acid and total NPC were the highest in the IA-II treatment. Quercetin 3-glucoside was significantly higher ($p \leq 0.05$) in the IA-I treatment in both years. Considering the other NPCs, values significantly increased in the non-irrigated vines ($p \leq 0.05$) (Table 6 and 7). Flavanols (Table 5), phenol acids (except caffeic acid) (Table 6), and total NPC values were significantly higher in the 2BL treatment ($p \leq 0.05$) compared to the 1BL treatment. The total NPC values ranged from 403.29 mg·kg⁻¹ and 449.94 mg·kg⁻¹ (in the non-irrigated control) to 486.34 mg·kg⁻¹ and 524.33 mg·kg⁻¹ (in the IA-II treatment) in 2013 and 2014, respectively (Table 7). These values are in accordance with results from Cantos et al. (2002) in which total phenolics ranged from 115 (Dominga) to 361 (Flame seedless) mg·kg⁻¹ fresh weight of berries. Lachman et al. (2009) also indicated that the values of

Table 7. Effects of irrigation amount (IA)[z] and bud load (BL) on flavonol, total non-colored phenolic compounds (NPC) and antioxidant contents of Kalecik Karası

Source of variance[y]	Rutin	Myricetin 3-glucoside	Isorhamnetin 3-glucoside	Quercetin 3-glucoside	Total NPC	Antioxidant activity
			(mg·kg⁻¹ fresh weight)			(mM Trolox·kg⁻¹)
			2013			
Water amounts						
No irrigation	1.22 a	6.22 a	1.53 a	2.83 b	403.29 c	289.61 a
IA-I	1.03 c	4.09 c	1.15 b	2.97 a	484.10 b	281.65 b
IA-II	1.15 b	5.21 b	0.97 c	2.37 c	486.34 a	287.54 a
(Pr > F)	***	***	***	***	***	**
Bud load						
1BL	1.19 a	5.52 a	1.37 a	2.95 a	421.38 b	284.83 b
2BL	1.07 b	4.82 b	1.06 b	2.50 b	494.44 a	287.71 a
(Pr > F)	***	***	***	***	***	*
Irr. × BL (Pr > F)	***	***	***	***	***	**
			2014			
Water amounts						
No irrigation	1.30 a	6.63 a	1.69 a	3.30 b	449.94 c	612.75 b
IA-I	1.10 c	4.36 c	1.27 b	3.46 a	519.88 b	549.50 c
IA-II	1.22 b	5.55 b	1.07 c	2.77 c	524.33 a	647.13 a
(Pr > F)	***	***	***	***	***	**
Bud load						
1BL	1.27 a	5.89 a	1.52 a	3.44 a	460.50 b	637.86 a
2BL	1.14 b	5.14 b	1.17 b	2.91 b	535.60 a	568.39 b
(Pr > F)	***	***	***	***	***	**
Irr. × BL (Pr > F)	***	***	***	***	***	**
Year (Pr > F)	***	***	***	***	***	***
Year × Irr. (Pr > F)	***	***	***	***	***	***
Year × BL (Pr > F)	***	***	***	***	***	***
Year × Irr. × BL (Pr > F)	***	***	***	***	***	***

[z]Values for the water amount main effects are the mean of 4 measurements (2 bud loads and 2 replicates) and values for the bud load main effects are the mean of 6 measurements (3 water amounts and 2 replicates).

[y]1BL (control) represents retaining 20 buds for the first 500 g of pruning weight plus 10 more buds for each additional 500 g., 2BL: retained double the buds of 1BL, irr.: Irrigation.

[x]Means followed by different letters on the same column are significantly different according to LSD test at $p \leq 0.05$.

[w]NS, *, ** or ***: Nonsignificant or significant at $p \leq 0.05$, 0.01, or 0.001, respectively.

total phenolics of berry skin obtained from different regions ranged from 370.1 mg·L⁻¹ to 238.7 mg·L⁻¹. Our results are also in accordance with the values found by Kelebek (2009), Göktürk Baydar et al. (2011), and Eshghi et al. (2014). Catechin and epicatechin are presumed to be the most important elements among the NPCs (Kennedy et al., 2002). According to Zarrouk et al. (2012), the effect of water amounts on grape flavanol levels seems to be controversial since they have been reported to be moderately affected by deficit irrigation and also not affected by water stress (Kennedy et al., 2002). In our study, the accumulation of total phenols and especially flavanols was enhanced by irrigation in comparison to the non-irrigated berries. This suggests that water availability has a positive effect on the accumulation and biosynthesis of phenols.

No clear relationship was found between the treatments and antioxidant contents (Table 7). The effects of the treatments on antioxidant content changed in the different years of this study. According to Kyraleou et al. (2015), the antioxidant

content of skin extracts was not affected by irrigation, whereas Özden et al. (2010) reported a decrease in antioxidants with increasing irrigation levels.

Romero et al. (2010) examined the effects of various levels of irrigation on phenolic compounds of Shiraz grapes during flowering through veraison and veraison through maturity growth stages. The biosynthesis of compounds varied according to irrigation levels and application periods. Similarly, other studies showed that fruit composition was slightly or inconsistently affected by water shortage and fruit load (Keller et al., 2008; Bowen et al., 2011).

Generally, water stress promotes the accumulation of sugar and anthocyanin (Roby et al. 2004; Castellarin et al., 2007) at the expense of a decrease in yield. Yield results of our study (data not shown) showed that an increase in yield can occur with moderate irrigation and increased bud loads in the high plateau regions without significant changes in the values of important health-related compounds.

Literature Cited

Beslic ZS, Todic SR, Tesevic VV, Jadranin MB, Novakovic MM, Tesic D (2010) Pruning effect on content of quercetin and catechin in berry skins of cv Blaufränkisch (*Vitis vinifera* L.). Turk J Agric For 34:461-466

Bindon K, Myburgh P, Oberholster A, Roux K, Du Toit C (2011) Response of grape and wine phenolic composition in *Vitis vinifera* L. cv. Merlot to variation in grapevine water status. S Afr J Enol Vitic 32:71-88

Bowen P, Bogdanoff C, Kevin Usher K, Estergaard B, Watson M (2011) Effects of irrigation and crop load on leaf gas exchange and fruit composition in red wine grapes grown on loamy sand. Am J Enol Viticult 62:9-22. doi:10.5344/ajev.2010.10046

Breksa AP, Takeoka GR, Hidalgo MB, Vilches A, Vasse J, Ramming DW (2010) Antioxidant activity and phenolic content of 16 raisin grape (*Vitis vinifera* L.) cultivars and selections. Food Chem 121:740-745. doi:10.1016/j.foodchem.2010.01.029

Bucchetti B, Matthews MA, Falginella L, Peterlunger E, Castellarin SD (2011) Effect of water deficit on Merlot grape tannins and anthocyanins across four seasons. Sci Hortic 128:297-305. doi:10.1016/j.scienta.2011.02.003

Cantos E, Espin JC, Tomas-Barberan FA (2002) Varietal differences among the polyphenol profiles of seven table grape cultivars studied by LC DAD MS MS. J Agric Food Chem 50:5691-5696. doi:10.1021/jf0204102

Castellarin S, Matthews M, Di Gaspero G, Gambetta G (2007) Water deficits accelerate ripening and induce changes in gene expression regulating flavonoid biosynthesis in grape berries. Planta 227:101-112. doi:10.1007/s00425-007-0598-8

Chalmers YM (2007) Influence of sustained deficit irrigation on physiology and phenolic compounds in winegrapes and wine. Available via https://digital.library.adelaide.edu.au/dspace/bitstream/2440/50101/8/02whole.pdf. Accessed 29 April 2016

Chaves MM, Santos TP, Souza CR, Ortuño MF, Rodrigues ML, Lopes CM, Maroco JP, Pereira JS (2007) Deficit irrigation in grapevine improves water use efficiency while controlling vigour and production quality. Ann Appl Biol 150:237-252. doi:10.1111/j.1744-7348.2006.00123.x

Cus F, Koruza ZK, Koruzo B, Lavrencic P (2004) Influence of crop load and cultivar's dependent canopy management on grape must quality of 'Sipon' 'Zametovka' and 'Rebula'. Acta Hortic 652:141-146. doi:10.17660/ActaHortic.2004.652.16

Çelik H, Ağaoğlu YS, Fidan Y, Marasalı B, Söylemezoğlu G (1998) General Viticulture (in Turkish). Sunfidan AŞ, Mesleki Kitaplar Serisi:1, Manisa, Turkey, pp 253

Çelik S (2011) Viticulture (Ampeloloja) Cilt-1 (in Turkish). Namık Kemal Univ Agric Fac Hortic Dept Tekirdag, Avcı Ofset İstanbul, Turkey, pp 428

Downey MO, Harvey JS, Robinson SP (2004) The effect of bunch shading on berry development and flavonoid accumulation in Shiraz grapes. Aust J Grape Wine R 10:55-73. doi:10.1111/j.1755-0238.2004.tb00008.x

Eshghi S, Salehi L, Karami MJ (2014) Antioxidant activity total phenolic compounds and anthocyanin contents in 35 different grapevine (*Vitis vinifera* L.) cultivars grown in Fars province. Int J Hortic Sci Technol 1:151-161

Esteban MA, Villanueva MJ, Lissarrague JR (2001) Effect of irrigation on changes in the anthocyanin composition of the skin of cv. Tempranillo (*Vitis vinifera* L.) grape berries during ripening. J Sci Food Agric 8:409-420. doi:10.1002/1097-0010(200103)81:4

Fawzi MIF, Shahin MFM, Kandil EA (2010) Effect of bud load on bud behaviour yield cluster characteristics and some biochemical contents of the cane of Crimson Seedless grapevines. J Am Sci 6:187-194

Göktürk Baydar N, Babalık Z, Hallaç Türk F, Çetin ES (2011) Phenolic composition and antioxidant activities of wines and extracts of some grape varieties grown in Turkey. J Agric Sci 17:67-76

Kallithraka S, Mohdaly AA, Makris DP, Kefalas P (2005) Determination of major anthocyanin pigments in Hellenic native grape varieties (*Vitis vinifera* sp.): Association with antiradical activity. J Food Compos Anal 18:375-386. doi:10.1016/j.jfca.2004.02.010

Kelebek H (2009) Researches on the phenolic compounds profile of Öküzgözü Bo azkere and Kalecik karası cultivars grown in different regions and their wines. Available via http://library.cu.edu.tr/tezler/7446.pdf. Accessed 29 April 2016

Kelebek H, Jourdes M, Selli S, Teissedre PL (2013) Comparative evaluation of the phenolic content and antioxidant capacity of sun-dried raisins. J Sci Food Agric 93:2963-2972

Keller M, Smithyman RP, Mills LJ (2008) Interactive effects of deficit irrigation and crop load on Cabernet Sauvignon in an arid climate. Am J Enol Viticult 59:221-234

Kennedy JA, Matthews MA, Waterhouse AL (2002) Effect of maturity and vine water status on grape skin and wine flavonoids. Am J Enol Viticult 53:268-274

Kurtural SK, Wessner LF, Dervishian G (2013) Vegetative compensation response of a procumbent grapevine (*Vitis vinifera* cv. Syrah) cultivar under mechanical canopy management. Hortscience 48:576-583

Kyraleou M, Koundouras S, Kallithraka S, Theodorou N, Proxenia N, Kotseridis Y (2015) Effect of irrigation regime on anthocyanin content and antioxidant activity of *Vitis vinifera* L. cv. Syrah grapes under semiarid conditions. J Sci Food Agric (online published). doi:101002/jsfa7175

Lachman, J, Šulc M, Faitová K, Pivec V (2009) Major factors influencing antioxidant contents and antioxidant activity in grapes and wines. Int J Wine Res 1:101-121. doi:10.2147/IJWR.S4600

Nelson CC, Kennedy JA, Zhang YI, Kurtural SK (2015) Applied water and rootstocks affect productivity and anthocyanin composition of Zinfandel in central California. Am J Enol Viticult (in press). doi:105344/ajev(2015)15043

Ozden M, Vardin H, Simsek M, Karaaslan M (2010) Effects of rootstocks and irrigation levels on grape quality of *Vitis vinifera* L. cv. Shiraz. Afr J Biotechnol 9:3801-3807

Rahmani M, Bakhshi D, Qolov M (2015) Impact of pruning severity and training systems on red and white seedless table grape (*Vitis vinifera*) qualitative indices. Aust J Crop Sci 9:55-61

Roby G, Matthews M (2004) Relative proportions of seed skin and flesh in ripe berries from Cabernet Sauvignon grapevines grown in a vineyard either well irrigated or under water deficit. Aust J Grape Wine R 10:74-82. doi:10.1111/j.1755-0238.2004.tb00009.x

Romero P, Fernandez-Fernandez JI, Martinez-Cutillas A (2010) Physiological thresholds for efficient regulated deficit-irrigation management in winegrapes grown under semi arid conditions. Am J Enol Viticult 61:300-312

Santesteban LG, Miranda C, Royo JB (2011) Regulated deficit irrigation effects on growth yield grape quality and individual anthocyanin composition in *Vitis vinifera* L. cv. 'Tempranillo'. Agric Water Manage 98:1171-1179. doi:10.1016/j.agwat.2011.02.011

Shellie KC (2011) Interactive effects of deficit irrigation and berry exposure aspect on Merlot and Cabernet Sauvignon in an arid climate. Am J Enol Viticult 62:462-470. doi:10.5344/ajev.2011.10103

Silva LR, Andrade PB, Valentao P, Seabra RM, Trujillo ME, Velazquez E (2005) Analysis of non-coloured phenolics in red wine: effect of *Dekkera bruxellensis* yeast. Food Chem 89:185-189. doi:10.1016/j.foodchem.2004.02.019

Sturm K, Koron D, Stampar F (2003) The Composition of fruit of different strawberry varieties depending on maturity stage. Food Chem 83:417-422. doi:10.1016/S0308-8146(03)00124-9

Sun J, Liang F, Bin Y, Li P, Duan C (2007) Screening non-colored phenolics in red wines using liquid chromatography/ultraviolet and mass spectrometry/mass spectrometry libraries. Molecules 12:679-693. doi:10.3390/12030679

Terry DB, Kurtural SK (2011) Achieving vine balance of Syrah with mechanical canopy management and regulated deficit irrigation. Am J Enol Viticult 62:426-437. doi:10.5344/ajev.2011.11022

Ward J (1984) The complete book of vine growing in the British Isles. Faber and Faber, London, Boston, pp 191

Weaver RJ (1976) Grape growing. Wiley Interscience Pub, pp 371

Wessner LF, Kurtural SK (2013) Pruning systems and canopy management practice interact on the yield and fruit composition of Syrah. Am J Enol Viticult 64:134-138. doi:10.5344/ajev.2012.12056

Wilson PE, Archbold DD, Masabni JG, Kurtural SK (2014) Cropload management of 'Vidal blanc' improves primary bud cold hardiness and maintains berry composition in the lower midwestern United States. Hortscience 49:874-880

Winkler AJ, Cook JA, Kliewer WM, Lider LA (1974) General viticulture. Univ. of California Press Berkeley, Los Angeles and London, pp 710

Zarrouk O, Francisco R, Pinto-Marijuan M, Brossa R, Santos RR, Pinheiro C, Costa JM, Lopes C, Chaves MM (2012) Impact of irrigation regime on berry development and flavonoids composition in Aragonez (Syn. Tempranillo) grapevine. Agric Water Manage 114:18-29. doi:10.1016/j.agwat.2012.06.018

Zhu L, Yali Z, Deng J, Li H, Lu J (2012) Phenolic concentrations and antioxidant properties of wines made from American grapes grown in China. Molecules 17:3304-3323. doi:10.3390/molecules17033304