

# Generation of Adventitious Roots and Characteristics of Gas Exchange according to Leaf Number of Hemp (*Cannabis sativa* L.) Cuttings

Vu Phong Lam<sup>1,2†</sup>, Ricardo Hernandez<sup>3†</sup>, Jeongyeo Lee<sup>4</sup>, Sung Jin Kim<sup>1</sup>, and Jongseok Park<sup>1\*</sup>

<sup>1</sup>Department of Horticultural Science, Chungnam National University, Daejeon 34134, Korea

<sup>2</sup>Department of Agronomy, Tay Bac University, Son La 360000, Vietnam

<sup>3</sup>Department of Horticultural Science, North Carolina State University, Raleigh 27695, USA

<sup>4</sup>Plant Systems Engineering Research Center, Korea Research Institute of Bioscience and Biotechnology, Daejeon 34141, Korea

\*Corresponding author: [jongseok@cnu.ac.kr](mailto:jongseok@cnu.ac.kr)

†These authors contributed equally to the work.

Received: June 1, 2021

Revised: November 10, 2021

Accepted: November 24, 2021

 OPEN ACCESS



HORTICULTURAL SCIENCE and TECHNOLOGY  
40(1):30-38, 2022  
URL: <http://www.hst-j.org>

pISSN : 1226-8763  
eISSN : 2465-8588

This is an Open Access article distributed under the terms of the Creative Commons Attribution Non-Commercial License which permits unrestricted non-commercial use, distribution, and reproduction in any medium, provided the original work is properly cited.

Copyright©2022 Korean Society for Horticultural Science.

This work was supported by research fund of Chungnam National University.

## Conflicts of Interest

Authors declare no conflict of interest.

## Author Contribution Statement

V.P.L. data collection and analysis, writing-original manuscript, writing-review and editing. R.H. experimental design, writing-review and editing. J.L. preparation for the manuscript. S.J.K. preparation for the manuscript. J.S.P. Project administration, supervision, constructing idea, experimental design, data analysis, writing-original manuscript, writing-review and editing.

## Abstract

In the hemp-seedling industry, the number of leaves on the harvested cuttings from mother plants is crucial for survival. Therefore, the present study determined the suitable number of leaves on hemp (*Cannabis sativa* L.) cuttings to promote adventitious root (AR) formation. Hemp cuttings with two, three, and four leaves were harvested from the apex of the mother plant. After disinfecting their base, 18 cuttings per replication were transplanted into a plug tray (72 holes) filled with a horticultural substrate. The photosynthetic characteristics of the cuttings and well-rooted hemp plants were measured at 22 and 60 days after transplantation (DAT), respectively. At 22 DAT, the average AR length, AR fresh weight, AR generation rate, and average number of ARs were the highest from two-leaf cuttings. Due to the higher AR generation, the net photosynthetic rate was the highest in cuttings with two-leaf cuttings at all the light intensity conditions (0, 50, 100, 175, 250, 500, 750, 1,000, and 1,500  $\mu\text{mol}\cdot\text{m}^{-2}\cdot\text{s}^{-1}$ ). The initial AR development of cuttings with two leaves likely enabled sufficient water uptake and photosynthesis to increase the survival rate. Growers may increase the productivity per unit area of well-rooted hemp plants by harvesting cuttings with two leaves.

**Additional key words:** fresh weight, light intensity, net photosynthetic rate, root quality, well-rooted hemp plant

## Introduction

In North America, cannabis (*Cannabis sativa* L.) production for legal markets, including the recreational and medicinal industries, is becoming profitable. North American spending on legal cannabis is estimated to reach \$22.6 billion in 2021 (ArcView Market Research, 2017). The medicinal value of cannabis is mainly attributed to a group of secondary metabolites called cannabinoids, which are mostly concentrated in unfertilized female flowers (Potter, 2014). Nowadays, some

countries including the United States, Canada, and the Netherlands have mitigated their legislations and established programs that permit the use of cannabis for medicinal purposes. In these programs, stringent safety criteria are enforced to manage the quality of cannabis administered to patients; however, growers are allowed a few manipulations for horticultural management (Caplan et al., 2017; Caplan et al., 2018). Currently, cannabis is mainly produced under controlled environments using a hydroponic system with artificial light (Caplan et al., 2017; Caplan et al., 2018). Hemp can be propagated by seeds (David, 2009), *in vitro* methods (Lata et al., 2009), and vegetative stem cuttings (Caplan et al., 2018). Hemp growers often prefer propagation by vegetative stem cuttings because they provide uniform plants with lower cost compared to propagation by seeds or *in vitro* methods (Caplan et al., 2018). The most important objective of propagation by vegetative stem cuttings is to promote adventitious roots (ARs) (Lebedev, 2019). Leaf number, cutting position on the plant, lighting, rooting hormones, water status, mineral nutrition, and rooting medium affect adventitious rooting in vegetative stem cuttings (Park et al., 2011; De Almeida et al., 2017; Park et al., 2017; Caplan et al., 2018). In this study, we focused on leaf numbers of hemp cuttings.

In some plant species, leaf retention on the stem cuttings is essential to ensure successful propagation (Hartmann and Kester, 1975; Park et al., 2011). Leaves are the major organ of photosynthesis for cuttings and are key to successful rooting. Increasing leaf number may enhance AR formation and rooting success rate in cuttings (Ofori et al., 1996; Caplan et al., 2018). Moreover, leaves can enhance rooting by serving as the source of endogenous auxin and rooting co-factors (Tombesi et al., 2015). In contrast, a larger leaf area and higher leaf number provide a greater surface area for evapotranspiration, which may negatively influence the rooting rate (Davis and Potter, 1989). The optimal leaf number on stem cuttings in lychee was four, although it varies among species (Alves et al., 2016).

Light intensity is the key regulator of photosynthesis, and it is involved at the photoreaction stage of photosynthesis by supplying sufficient electrons for dark responses (Zhou et al., 2012; Zhang et al., 2020). Plant responses to high CO<sub>2</sub> concentrations are mediated by leaf photosynthesis, which is associated with changes in leaf structure, carbon balance, and chemical composition, depending on the plant species (Lee et al., 2001; Ainsworth et al., 2002; Zheng et al., 2019b). Photosynthetic characteristics at the rooting stage of cuttings have been examined in some plant species, such as poinsettia (Svenson et al., 1995), *Acer rubrum* (Smalley et al., 1991), and petunia (Klopotek et al., 2012), but not in hemp. Therefore, the objective of the present study was to explore the effects of leaf number on hemp cuttings, grown for medicinal purposes on AR generation and to understand the photosynthetic characteristics of this species.

## Materials and Methods

### Stock Plant Conditions

Hemp (*Cannabis sativa* L.) mother plants were cultivated in a greenhouse controlled at 25/18°C ± 2°C (day/night) with a pad and fan cooling system at North Carolina State University. The mother plants were grown in pots (approximately 30 L) containing a blended organic growing substrate and watered 3–5 times a day using an automated drip irrigation system with Hoagland nutrient solution (1.2 dS·m<sup>-1</sup> electrical conductivity and 5.8 pH) for 8 months (Lam et al., 2020).

## Plant Culture Treatments and Propagation Environment

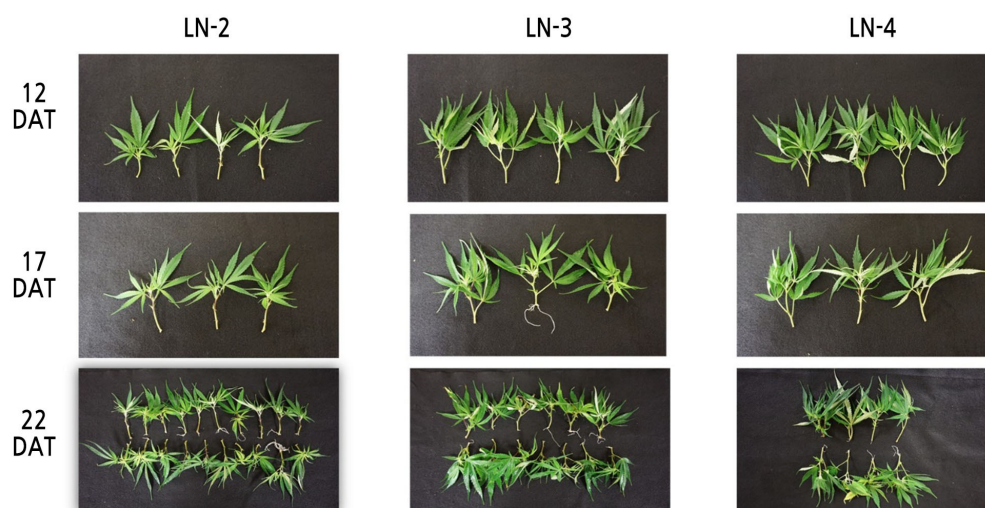
Approximately 150 hemp cuttings with two (two fully expanded top leaves), three (three fully expanded top leaves), and four leaves (four fully expanded top leaves) were harvested from the apical regions of 5 mother plants, and 18 uniform cuttings per replication were selected (Fig. 1). We defined the leaf with a leaf length of three cm or more. We conducted two replications per treatment. The cuttings were dipped into 1% diluted soap water and then into a bactericide (ZeroTol 2.0, Biosafe System, LLC., East Hartford, CT, USA) diluted to 37% (0.5 ozone/1 gallon) for one minute for disinfection and transplanted in a plug tray (72 holes) filled with a horticultural substrate (Fafard 4P Mix, Sun Gro Horticulture Co. Ltd, Agawam, MA, USA). The trays were randomly placed in an indoor cultivation room controlled at  $25^{\circ}\text{C} \pm 2^{\circ}\text{C}$  average temperature with 95%, 90%, and 85% relative humidity in the first, second, and third weeks, respectively. Red and blue LEDs were installed for light irradiation at approximately  $80 \mu\text{mol}\cdot\text{m}^{-2}\cdot\text{s}^{-1}$  with a photoperiod of 16 h for 22 days after transplantation (DAT). Cuttings were watered with an ebb and flow system every 5 min for 22 DAT.

## Rooting Assessment

The tray bottom was checked daily from 12 DAT onward for ARs, and cuttings were harvested at 22 DAT. The substrate was removed from the rooted cuttings by washing with water. The average length of ARs was determined with a measuring tape (STHT-36127, Stanley Electric Co., Ltd., Tianjin, China), and the average fresh weight of ARs was measured using an electronic scale (Si-234, Denver Instrument, Bohemia, NY, USA). The average AR number was also counted. The AR generation rate was calculated as the percentage of rooted cuttings in each treatment. All AR characteristics were determined at 22 DAT on eighteen cuttings per replication ( $n = 18$ ).

## Photosynthetic Characteristics Measurements

A portable photosynthesis system (LI-6800XT, LI-COR Biosciences Inc., Lincoln, NE, USA) was used to measure the net photosynthetic rate, stomatal conductance, and transpiration rate of cuttings. Five plants ( $n = 5$ ) in each replication



**Fig. 1.** Photographs of cannabis cuttings with two (LN-2), three (LN-3), and four (LN-4) leaves at 12, 17, and 22 days after transplantation (DAT).

were used and the measurement was performed with two replications per treatment. The net photosynthetic rate of cuttings with two, three, and four leaves was measured at 22 DAT and that of well-rooted hemp plants was measured at 60 DAT. For measuring the net photosynthetic rate of cuttings with two, three, and four leaves at 22 DAT, the light intensity (photosynthetic photon flux density [PPFD]) was set at 0, 50, 100, 175, 250, 500, 750, 1,000, and 1,500  $\mu\text{mol}\cdot\text{m}^{-2}\cdot\text{s}^{-1}$ ;  $\text{CO}_2$  concentration was set at 400  $\mu\text{mol}\cdot\text{mol}^{-1}$ , and chamber temperature was set at 25°C. In measurement 1 at 60 DAT, photosynthetic characteristics were measured at PPFD of 0, 50, 100, 175, 250, 500, 750, 1,000, and 1,500  $\mu\text{mol}\cdot\text{m}^{-2}\cdot\text{s}^{-1}$ ; chamber temperature of 25°C; and  $\text{CO}_2$  concentration of 400  $\mu\text{mol}\cdot\text{mol}^{-1}$ . In measurement 2 at 60 DAT, photosynthetic characteristics were measured at  $\text{CO}_2$  concentrations of 50, 100, 200, 300, 400, 600, 800, 1,000, and 1,500  $\mu\text{mol}\cdot\text{mol}^{-1}$ ; PPFD of 1,500  $\mu\text{mol}\cdot\text{m}^{-2}\cdot\text{s}^{-1}$ ; and chamber temperature of 25°C.

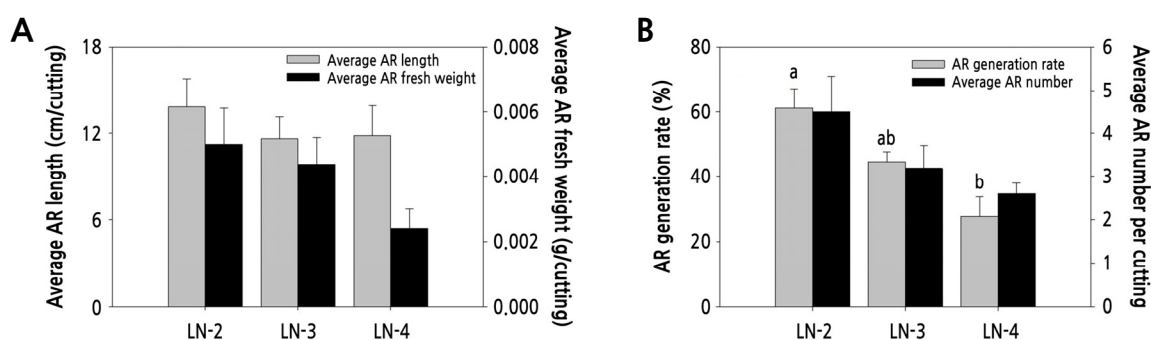
## Statistical Analysis

In this experiment, we conducted two replications by using completely randomized designs. The SPSS 20.0 program (SPSS 20, SPSS Inc., Chicago, IL, USA) was used for a one-way analysis of variance. Significant differences among the means of treatment groups at the 5% level were compared using Tukey's multiple range test.

## Results and Discussion

### Adventitious Root Development

Adventitious root (AR) development in hemp cuttings was affected by the number of leaves. At 22 DAT, there was no significant difference among leaf number treatments; however, the two-leaf treatment showed higher values of average AR length, average AR fresh weight, AR generation rate, and average number of ARs compared with other treatments (Figs. 1 and 2). Two leaves per cutting likely reduced evapotranspiration; therefore, fewer leaves would enhance root growth because of reduced evapotranspiration. Previous studies have shown the effects of cutting size and leaf number as well as of cutting length  $\times$  leaf area interaction on rooting (Tchoundjeu and Leakey, 1996; OuYang et al., 2015). The number of mature leaves on cuttings at the time of transplantation is important for the production of new leaves (Yamdagni and Sen, 1973). Root weight, root length, and roots number were affected by leaflets left on each cutting (Park



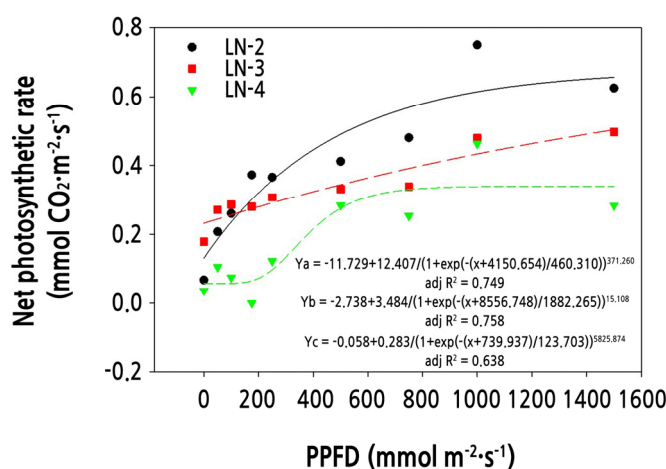
**Fig. 2.** Average adventitious root (AR) length (A left), average AR fresh weight (A right), AR generation rate (B left), and average AR numbers (B right) in hemp cuttings with two (LN-2), three (LN-3), and four (LN-4) leaves. The cuttings were obtained from the apical regions of the mother plant. Values are presented as mean  $\pm$  SE ( $n = 18$ ).

et al., 2011). Leaves may directly affect shoot growth since the initial growth of shoots in seedlings depends on the carbon assimilation by mature leaves (Costa and Challa, 2002). Leaves on cuttings support CO<sub>2</sub> assimilation throughout the rooting period (Smalley et al., 1991; Klopotek et al., 2012). At the initial stages of cutting growth, water uptake occurs partially at the base of the cut surface in the absence of ARs. The higher transpiration rate of cuttings with four leaves than that of cuttings with two leaves may lead to physiological disabilities (Newton et al., 1992). Typically, at the early stages of rooting, high humidity is maintained to inhibit transpiration and induce AR formation (De Almeida et al., 2017). Our results indicate that cuttings with two leaves are the most suitable for the adventitious rooting of hemp.

### Photosynthetic Characteristics of Hemp Cuttings with Different Leaf Numbers

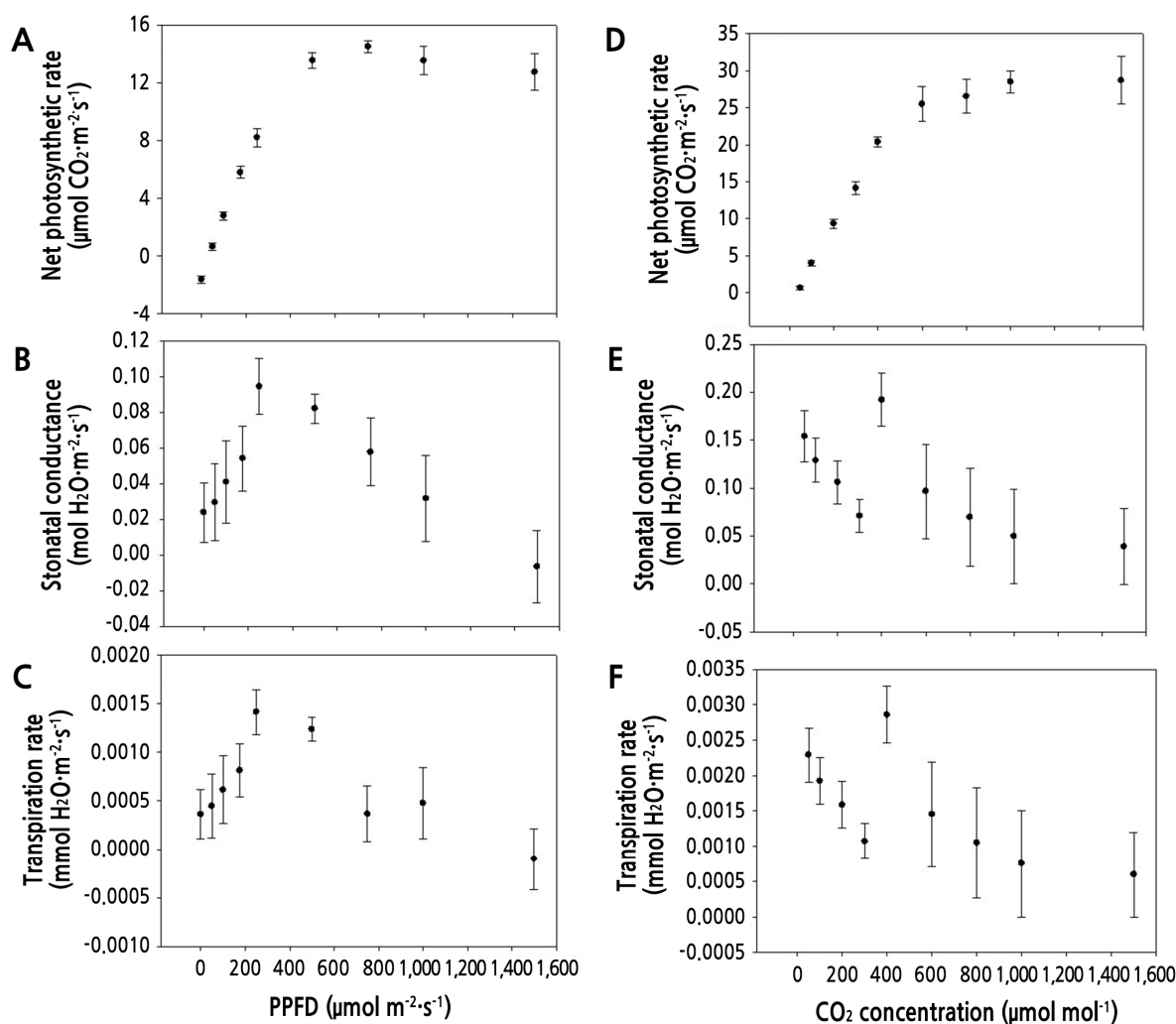
The net photosynthesis rate of hemp cuttings with different leaf numbers was measured under various light intensities and a constant CO<sub>2</sub> concentration and temperature (400  $\mu\text{mol}\cdot\text{mol}^{-1}$  and 25°C, respectively) (Fig. 3). There were significant differences in the net photosynthetic rate of cuttings with different leaf numbers. Cuttings with two leaves showed the highest net photosynthetic rate probably because of reduced water loss from fewer leaves. Although cuttings with two and three leaves showed similar responses to different light intensities, the two-leaf cuttings achieved a higher net photosynthetic rate than the three-leaf cuttings. These results indicate that cuttings with two leaves were more efficient in initial AR formation and carbohydrate supply to these ARs than cuttings with three leaves. At 22 DAT, the net photosynthetic rate was the highest in cuttings with two leaves probably due to earlier AR development.

Photosynthetic characteristics of hemp were observed under various light intensities and influenced by changing the amount of light intensity. The received light is weak when the light intensity is low; therefore, the plant will create more photosynthetic pigments, especially chlorophyll (Chl) b, resulting in a decrease of the Chl a/b ratio as well as a decrease of the photosynthetic characteristics. At 60 DAT, the net photosynthetic rate of hemp markedly increased with increasing light intensity from 0 to 750  $\mu\text{mol}\cdot\text{m}^{-2}\cdot\text{s}^{-1}$ . However, the net photosynthetic rate became saturated from 760 to 1,500  $\mu\text{mol}\cdot\text{m}^{-2}\cdot\text{s}^{-1}$  (Fig. 4A). The stomatal conductance of cuttings increased with increasing light intensity from 0 to 250



**Fig. 3.** Regression analyses of mean photosynthesis rate on two (LN-2), three (LN-3), and four (LN-4) leaves for cuttings at 22 days after transplantation under different light intensities (photosynthetic photon flux density, PPFD) (0, 50, 100, 175, 250, 500, 750, 1,000, and 1,500  $\mu\text{mol}\cdot\text{m}^{-2}\cdot\text{s}^{-1}$ ). The solid, long dashed, and short dashed lines in the graph represent the quadratic regression for two (LN-2), three (LN-3), and four (LN-4) leaves and the relation equation is given as  $Y_a$ ,  $Y_b$ , and  $Y_c$ , respectively.

$\mu\text{mol}\cdot\text{m}^{-2}\cdot\text{s}^{-1}$  and then decreased with increasing light intensity from 300 to  $1,500\ \mu\text{mol}\cdot\text{m}^{-2}\cdot\text{s}^{-1}$  (Fig. 4B). Likewise, the transpiration rate of hemp plants increased with increasing light intensity from 0 to  $250\ \mu\text{mol}\cdot\text{m}^{-2}\cdot\text{s}^{-1}$  and then decreased with increasing light intensity from 300 to  $1,500\ \mu\text{mol}\cdot\text{m}^{-2}\cdot\text{s}^{-1}$  (Fig. 4C). These results corroborate the findings reported by Zheng et al. (2019b); the authors found that the net photosynthetic rate markedly increased from 0 to  $600\ \mu\text{mol}\cdot\text{m}^{-2}\cdot\text{s}^{-1}$  but remained stable from 800 and  $1,800\ \mu\text{mol}\cdot\text{m}^{-2}\cdot\text{s}^{-1}$  (Zheng et al., 2019a). Thus, based on the net photosynthetic rate, it is economical to cultivate rooted hemp plants at a light intensity of  $\sim 600\ \mu\text{mol}\cdot\text{m}^{-2}\cdot\text{s}^{-1}$ . Well-rooted hemp cuttings are adversely affected by high light intensity because of increased leaf transpiration and temperature (Zobolo, 2010). Leaf photosynthetic activity rapidly increases with increasing light intensity from 0 to  $500\ \mu\text{mol}\cdot\text{m}^{-2}\cdot\text{s}^{-1}$ , probably due to an increase in the levels of non-structural carbohydrates (Tombesi et al., 2015). Light intensity levels below the saturation peak can promote the photosynthetic activity of cuttings; however, extreme light regimes can enhance the air vapor pressure deficit, resulting in leaf desiccation (Tombesi et al., 2015; Zheng et al., 2019a). Until the saturation of the net



**Fig. 4.** Net photosynthetic rate (A and D), stomatal conductance (B and E), and transpiration rate (C and F) of hemp at 60 days after transplantation under different light intensities (0, 50, 100, 175, 250, 500, 750, 1,000, and  $1,500\ \mu\text{mol}\cdot\text{m}^{-2}\cdot\text{s}^{-1}$ ) and different  $\text{CO}_2$  concentrations (300, 200, 100, 50, 400, 600, 800, 1,000, and  $1,500\ \mu\text{mol}\cdot\text{mol}^{-1}$ ). Values are presented as mean  $\pm$  SE ( $n = 5$ ).



photosynthetic rate, the stomatal conductance, and transpiration rate tend to increase. However, when the photosynthetic saturation point is reached, the plants gradually close the stomata to control transpiration while maintaining the photosynthetic saturation state and accelerating the CO<sub>2</sub> exchange rate. The light saturation point of hemp is not as high as that of other horticultural crops (Xin et al., 2019); therefore, a light intensity over 400  $\mu\text{mol}\cdot\text{m}^{-2}\cdot\text{s}^{-1}$  should be set to promote hemp rooting in an indoor cultivation system.

### Photosynthetic Characteristics of Hemp Under Various CO<sub>2</sub> Concentrations

The net photosynthetic rate of cuttings increased markedly with increasing CO<sub>2</sub> concentration from 50 to 600  $\mu\text{mol}\cdot\text{mol}^{-1}$  and became saturated above the concentration of 600  $\mu\text{mol}\cdot\text{mol}^{-1}$  (Fig. 4D). The stomatal conductance and transpiration rate increased in the order of initial CO<sub>2</sub> concentrations of 300, 200, 100, 50, and 400  $\mu\text{mol}\cdot\text{mol}^{-1}$  and then gradually decreased from 600 to 1,500  $\mu\text{mol}\cdot\text{mol}^{-1}$  (Fig. 4E and 4F). When the CO<sub>2</sub> concentration increases, the stomata close, and when the CO<sub>2</sub> concentration decreases, the stomata are ordinarily open. To analyze the gas exchange characteristics according to the CO<sub>2</sub> concentration, the order of the CO<sub>2</sub> setpoints was 300, 200, 100, 50, 400, 600, 800, 1,000, and 1,500  $\mu\text{mol}\cdot\text{mol}^{-1}$ . This is to confirm the pattern of increasing the stomatal conductance with decreasing the CO<sub>2</sub> concentration (from 300 to 50), and to confirm the tendency of the stomatal conductance to decrease as the CO<sub>2</sub> concentration increased under the condition of 400–1,500  $\mu\text{mol}\cdot\text{mol}^{-1}$ . Moreover, the CO<sub>2</sub> saturation point was determined to be 600  $\mu\text{mol}\cdot\text{mol}^{-1}$  in Fig. 4; however, the stomatal conductance reached the maximum value at 400  $\mu\text{mol}\cdot\text{mol}^{-1}$  CO<sub>2</sub> concentration point before reaching the CO<sub>2</sub> saturation point. These results indicate that elevated CO<sub>2</sub> levels increased the net photosynthetic rate of plants, particularly C<sub>3</sub> species (Singh and Reddy, 2016; Zheng et al., 2019b). The photosynthetic rate of the lettuce at 4, 7, 14, 21, and 28 days after transplanting was increased with increasing CO<sub>2</sub> concentration from 100 to 600  $\mu\text{mol}\cdot\text{mol}^{-1}$  (Jung et al., 2016). This increase in photosynthesis under elevated CO<sub>2</sub> concentrations may reduce dark respiration and photorespiration and enhance carboxylation efficiency (Duarte et al., 2013). Elevated CO<sub>2</sub> levels may increase photosynthetic carbon assimilation and potentially improve plant growth and productivity. However, the net photosynthetic rate remained unchanged when the plants were grown under long-term high CO<sub>2</sub> concentrations (Casteel et al., 2008; Robredo et al., 2010). In soybean, leaf photosynthesis was reduced with increasing CO<sub>2</sub> concentrations from 400 to 1,000  $\mu\text{mol}\cdot\text{mol}^{-1}$  (Kanemoto et al., 2009). This reduction in the net photosynthetic rate under high CO<sub>2</sub> concentrations may be attributed to a decline in Rubisco activity or carbohydrate accumulation in leaves (Jifon and Wolfe, 2002; Davey et al., 2006; Zheng et al., 2019b). Moreover, the suppression of photosynthesis at high CO<sub>2</sub> concentrations may be caused by reduced stomatal conductance and respiration (Hamilton et al., 2001; Wang et al., 2013). Leaf biochemical characteristics and structure may play important roles in plant responses to elevated CO<sub>2</sub> concentrations. Typically, high CO<sub>2</sub> concentrations increase leaf thickness and mesophyll size because of elevated carbohydrate substrate availability, which is related to leaf photosynthetic rate (Tissue and Lewis, 2010; Zheng et al., 2019b). In addition, high CO<sub>2</sub> concentrations might alter leaf biochemical composition, including the concentration of nitrogen and non-structural carbohydrates, which play essential roles in plant responses to elevated CO<sub>2</sub> (Xu et al., 2012). Therefore, to achieve the maximum net photosynthetic rate of hemp, the CO<sub>2</sub> concentration should be set to ~600  $\mu\text{mol}\cdot\text{mol}^{-1}$  with other suitable environmental conditions in an indoor cultivation system.

## Conclusion

The present study demonstrated that to obtain the maximum adventitious rooting in hemp cuttings derived from the apical region of the mother plant, the cuttings should bear two expanded leaves. Moreover, to achieve the maximum net photosynthetic rate in hemp, a PPFD of 500 to 750  $\mu\text{mol}\cdot\text{m}^{-2}\cdot\text{s}^{-1}$  and a  $\text{CO}_2$  concentration of 600  $\mu\text{mol}\cdot\text{mol}^{-1}$  may be the most suitable settings in an indoor cultivation system.

## Literature Cited

- Ainsworth EA, Davey PA, Bernacchi CJ, Dermody OC, Heaton EA, Moore DJ, Morgan PB, Naidu SL, Ra HSY, et al (2002) A meta-analysis of elevated  $[\text{CO}_2]$  effects on soybean (*Glycine max*) physiology, growth and yield. *Glob Chang Biol* 8:695-709. doi:10.1046/j.1365-2486.2002.00498.x
- Alves EC, Guimaraes JER, Franco CKB, Martins ABG (2016) Number of leaflets on rooting of lychee herbaceous cuttings. *Ciencia Rural* 46:1003-1006. doi:10.1590/0103-8478cr20140435
- ArcView Market Research (2017) The state of legal marijuana markets. 5th ed. ArcView Market Research, San Francisco, CA, USA
- Caplan D, Dixon M, Zheng YB (2017) Optimal rate of organic fertilizer during the flowering stage for cannabis grown in two coir-based substrates. *Hortscience* 52:1796-1803. doi:10.21273/HORTSCI12401-17
- Caplan D, Stemeroff J, Dixon M, Zheng YB (2018) Vegetative propagation of cannabis by stem cuttings: effects of leaf number, cutting position, rooting hormone, and leaf tip removal. *Canadian Journal of Plant Science* 98:1126-1132. doi:10.1139/cjps-2018-0038
- Casteel CL, O'Neill BF, Zavala JA, Bilgin DD, Berenbaum MR, Delucia EH (2008) Transcriptional profiling reveals elevated  $\text{CO}_2$  and elevated  $\text{O}_3$  alter resistance of soybean (*Glycine max*) to Japanese beetles (*Popillia japonica*). *Plant Cell Environ* 31:419-34. doi:10.1111/j.1365-3040.2008.01782.x
- Costa JM, Challa H (2002) The effect of the original leaf area on growth of softwood cuttings and planting material of rose. *Scientia Horticulturae* 95:111-121. doi:10.1016/S0304-4238(02)00023-7
- Davey PA, Olcer H, Zakhleniuk O, Bernacchi CJ, Calfapietra C, Long SP, Raines CA (2006) Can fast-growing plantation trees escape biochemical down-regulation of photosynthesis when grown throughout their complete production cycle in the open air under elevated carbon dioxide? *Plant Cell and Environment* 29:1235-1244. doi:10.1111/j.1365-3040.2006.01503.x
- David J (2009) The propagation, characterisation and optimisation of *Cannabis sativa* L. as a phytopharmaceutical. Ph.D. thesis, King's College London, London, UK
- Davis TD, Potter JR (1989) Relations between carbohydrate, water status and adventitious root-formation in feafy pea cuttings rooted under various levels of atmospheric  $\text{CO}_2$  and relative-humidity. *Physiol Plant* 77:185-190. doi:10.1111/j.1399-3054.1989.tb04967.x
- De Almeida MR, Aumond M, Da Costa CT, Schwambach J, Ruedell CM, Correa LR, Fett-Neto AG (2017) Environmental control of adventitious rooting in *Eucalyptus* and *Populus* cuttings. *Trees-Structure and Function* 31:1377-1390. doi:10.1007/s00468-017-1550-6
- Duarte B, Santos D, Marques JC, Cacador I (2013) Ecophysiological adaptations of two halophytes to salt stress: Photosynthesis, PS II photochemistry and anti-oxidant feedback - Implications for resilience in climate change. *Plant Physiol Biochem* 67:178-188. doi:10.1016/j.plaphy.2013.03.004
- Hamilton JG, Thomas RB, Delucia EH (2001) Direct and indirect effects of elevated  $\text{CO}_2$  on leaf respiration in a forest ecosystem. *Plant Cell and Environment* 24:975-982. doi:10.1046/j.0016-8025.2001.00730.x
- Hartmann HT, Kester DE (1975) Plant propagation: principles and practices. 3rd edition, Prentice-Hall, New Jersey, USA, p 662
- Jifon JL, Wolfe DW (2002) Photosynthetic acclimation to elevated  $\text{CO}_2$  in *Phaseolus vulgaris* L. is altered by growth response to nitrogen supply. *Glob Chang Biol* 8:1018-1027. doi:10.1046/j.1365-2486.2002.00531.x
- Jung DH, Kim D, Yoon HI, Moon TW, Park KS, Son JE (2016) Modeling the canopy photosynthetic rate of romaine lettuce (*Lactuca sativa* L.) grown in a plant factory at varying  $\text{CO}_2$  concentrations and growth stages. *Hortic Environ Biotechnol* 57:487-492. doi:10.1007/s13580-016-0103-z
- Kanemoto K, Yamashita Y, Ozawa T, Imanishi N, Nguyen NT, Suwa R, Mohapatra PK, Kanai S, Moghaieb RE, et al (2009) Photosynthetic acclimation to elevated  $\text{CO}_2$  is dependent on N partitioning and transpiration in soybean. *Plant Science* 177:398-403. doi:10.1016/j.plantsci.2009.06.017
- Klopotek Y, George E, Druege U, Klaering HP (2012) Carbon assimilation of petunia cuttings in a non-disturbed rooting environment: Response to environmental key factors and adventitious root formation. *Sci Hortic* 145:118-126. doi:10.1016/j.scienta.2012.08.004
- Lam VP, Kim SJ, Park JS (2020) Optimizing the electrical conductivity of a nutrient solution for plant growth and bioactive compounds of *Agastache rugosa* in a plant factory. *Agronomy* 10:76. doi:10.3390/agronomy10010076
- Lata H, Chandra S, Khan IA, Elsohly MA (2009) Propagation through alginate encapsulation of axillary buds of *Cannabis sativa* L. - an important medicinal plant. *Physiol Mol Biol Plants* 15:79-86. doi:10.1007/s12298-009-0008-8
- Lebedev V (2019) The rooting of stem cuttings and the stability of uidA gene expression in generative and vegetative progeny of



- transgenic pear rootstock in the field. *Plants-Basel* 8. doi:10.3390/plants8080291
- Lee TD, Tjoelker MG, Ellsworth DS, Reich PB** (2001) Leaf gas exchange responses of 13 prairie grassland species to elevated CO<sub>2</sub> and increased nitrogen supply. *New Phytologist* 150:405-418. doi:10.1046/j.1469-8137.2001.00095.x
- Newton AC, Muthoka PN, Dick JM** (1992) The influence of leaf area on the rooting physiology of leafy stem cuttings of *Terminalia spinosa* Engl. *Trees-Structure and Function* 6:210-215. doi:10.1007/BF00224338
- Ofori DA, Newton AC, Leakey RRB, Grace J** (1996) Vegetative propagation of *Milicia excelsa* by leafy stem cuttings: Effects of auxin concentration, leaf area and rooting medium. *For Ecol Manag* 84:39-48. doi:10.1016/0378-1127(96)03737-1
- OuYang FQ, Wang JH, Li Y** (2015) Effects of cutting size and exogenous hormone treatment on rooting of shoot cuttings in Norway spruce [*Picea abies* (L.) Karst.]. *New Forests* 46:91-105. doi:10.1007/s11056-014-9449-1
- Park SM, Won EJ, Park YG, Jeong BR** (2011) Effects of node position, number of leaflets left, and light intensity during cutting propagation on rooting and subsequent growth of domestic roses. *Hortic. Environ. Biotechnol* 53:339-343. doi:10.1007/s13580-011-0163-z
- Park SW, Kwack Y, Chun C** (2017) Growth of runner plants grown in a plant factory as affected by light intensity and container volume. *Hortic Sci Technol* 35:439-445. doi:10.12972/kjhst.20170047
- Potter DJ** (2014) A review of the cultivation and processing of cannabis (*Cannabis sativa* L.) for production of prescription medicines in the UK. *Drug Test Anal* 6:31-38. doi:10.1002/dta.1531
- Robredo A, Perez-Lopez U, Lacuesta M, Mena-Petite A, Munoz-Rueda A** (2010) Influence of water stress on photosynthetic characteristics in barley plants under ambient and elevated CO<sub>2</sub> concentrations. *Biol Plant* 54:285-292. doi:10.1007/s10535-010-0050-y
- Singh SK, Reddy VR** (2016) Methods of mesophyll conductance estimation: its impact on key biochemical parameters and photosynthetic limitations in phosphorus-stressed soybean across CO<sub>2</sub>. *Physiol Plant* 157:234-254. doi:10.1111/ppl.12415
- Smalley TJ, Dirr MA, Armitage AM, Wood BW, Teskey RO, Severson RF** (1991) Photosynthesis and leaf water, carbohydrate, and hormone status during rooting of stem cuttings of *Acer-rubrum*. *J Am Soc Hortic Sci* 116:1052-1057. doi:10.21273/JASHS.116.6.1052
- Svenson SE, Davies FT, Duray SA** (1995) Gas-exchange, water relations, and dry-weight partitioning during root initiation and development of poinsettia cuttings. *J Am Soc Hortic Sci* 120:454-459. doi:10.21273/JASHS.120.3.454
- Tchoundjeu Z, Leakey RRB** (1996) Vegetative propagation of African Mahogany: Effects of auxin, node position, leaf area and cutting length. *New Forests* 11:125-136. doi:10.1007/BF00033408
- Tissue DT, Lewis JD** (2010) Photosynthetic responses of cottonwood seedlings grown in glacial through future atmospheric [CO<sub>2</sub>] vary with phosphorus supply. *Tree Physiology* 30:1361-1372. doi:10.1093/treephys/tpq077
- Tombesi S, Palliotti A, Poni S, Farinelli D** (2015) Influence of light and shoot development stage on leaf photosynthesis and carbohydrate status during the adventitious root formation in cuttings of *Corylus avellana* L. *Front Plant Sci* 6:973. doi:10.3389/fpls.2015.00973
- Wang L, Feng ZZ, Schjoerring JK** (2013) Effects of elevated atmospheric CO<sub>2</sub> on physiology and yield of wheat (*Triticum aestivum* L.): A meta-analytic test of current hypotheses. *Agric Ecosyst Environ* 178:57-63. doi:10.1016/j.agee.2013.06.013
- Xin PP, Li B, Zhang HH, Hui J** (2019) Optimization and control of the light environment for greenhouse crop production. *Sci Rep* 9:8650. doi:10.1038/s41598-019-44980-z
- Xu CY, Salih A, Ghannoum O, Tissue DT** (2012) Leaf structural characteristics are less important than leaf chemical properties in determining the response of leaf mass per area and photosynthesis of *Eucalyptus saligna* to industrial-age changes in [CO<sub>2</sub>] and temperature. *J Exp Bot* 63:5829-5841. doi:10.1093/jxb/ers231
- Yamdagni N, Sen D** (1973) Role of leaves present on the stem cuttings for vegetative propagation in *Portulaca grandiflora* L. *Biochem Physiol Pflanz* 164:447-449. doi:10.1016/S0015-3796(17)30715-1
- Zhang P, Zhang ZX, Li B, Zhang HH, Hu J, Zhao J** (2020) Photosynthetic rate prediction model of newborn leaves verified by core fluorescence parameters. *Sci Rep* 10:3013. doi:10.1038/s41598-020-59741-6
- Zheng JF, Ji F, He DX, Niu GH** (2019a) Effect of light intensity on rooting and growth of hydroponic strawberry runner plants in a led plant factory. *Agronomy* 9:875. doi:10.3390/agronomy9120875
- Zheng YP, Li F, Hao LH, Yu JJ, Guo LL, Zhou HR, Ma C, Zhang XX, Xu M** (2019b) Elevated CO<sub>2</sub> concentration induces photosynthetic down-regulation with changes in leaf structure, non-structural carbohydrates and nitrogen content of soybean. *BMC Plant Biol* 19. doi:10.1186/s12870-019-1788-9
- Zhou WL, Liu WK, Yang QC** (2012) Quality changes in hydroponic lettuce grown under pre-harvest short-duration continuous light of different intensities. *J Hortic Sci Biotechnol* 87:429-434. doi:10.1080/14620316.2012.11512890
- Zobolo AM** (2010) Effect of temperature, light intensity and growth regulators on propagation of *Ansellia Africana* from cuttings. *Afr J Biotechnol* 9:5566-5574. doi:10.5897/AJB09.1495