

Analysis of Structure and Expression of Grapevine 2-oxoglutarate Oxygenase Genes in Response to Low Temperature

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Abstract

2-Oxoglutarate (2OG) acts as a signaling molecule and plays a critical role in secondary metabolism in a variety of organisms, including plants. Six 2-oxoglutarate (2OG) and Fe(II) oxygenase (2OGO) genes, *VICE2OGO1* [*Vitis labruscana* 2-oxoglutarate (2OG) and Fe(II) oxygenase 1], *VICE2OGO2*, *VICE2OGO3*, *VICE2OGO4*, *VICE2OGO5*, and *VICE2OGO6*, which show different expression patterns upon transcriptome analysis of ‘Campbell Early’ grapevine exposed to low temperature for 4 weeks, were analyzed for their structure and expression. Comparison of the deduced amino acid sequences of the 2OGO genes from the *V. labruscana* transcripts revealed sequence similarities of 38.6% (*VICE2OGO1* and *VICE2OGO2*) to 19.2% (*VICE2OGO2* and *VICE2OGO3*). The lengths of these genes ranged from 1053 to 2298 bp, and they encoded 316 to 380 amino acids. The prediction of the secondary structure of the encoded proteins by Self-Optimized Prediction Method with Alignment (SOPMA) indicated that all the genes contained alpha helix (23.95 to 41.71%), extended strand (16 to 22.34%), beta turn (6.65 to 9.22%), and random coil (32.97 to 51.58%) in the analysis. Specific primers from unique regions in each gene obtained by alignment of nucleotide sequences were used in real time PCR for analysis of gene expression. All tested genes showed differential expression in grapevines exposed to low temperature. Of the six transcripts, *VICE2OGO1*, *VICE2OGO2*, and *VICE2OGO3* were up-regulated and *VICE2OGO4*, *VICE2OGO5*, and *VICE2OGO6* were down-regulated in response to cold treatments at all tested time points. The 2OG genes can be used for elucidation of mechanisms of tolerance to cold and as valuable molecular genetic resources for selection in breeding programs for cold-hardy grapevines.

Additional key words: differential expression, tolerance to cold, *Vitis labruscana*

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Introduction

Temperature is one of primary environmental factors that limit the growth, development, and productivity of crop plants. Molecular and genomic analyses have identified several regulatory systems controlling expression of genes in response to stress (Shinozaki et al., 2003). Grape (*Vitis* spp.), which is one of the most widely grown fruit crops in the world, is also influenced by low temperature during the growing season (Nam et al., 2012; Park et al., 2000; Warmund et al., 2008). Tattersall et al. (2007)

conducted a microarray analysis of *V. vinifera* cv. Cabernet Sauvignon that found that chilling affects many calcium signaling-related transcripts in shoot tips of grapevines exposed to chilling.

2-Oxoglutarate (2OG) is a molecule derived from the Krebs cycle, which is a highly conserved metabolic process. This compound plays a critical role in secondary metabolism as a signaling molecule in a variety of organisms (Zhao et al., 2010). 2OG and Fe(II)-dependent oxygenases (2OGO) are the largest known family of non-heme oxidizing enzymes, which have an absolute requirement for desaturation, hydroxylation, and oxidative ring closure reactions (Prescott and John, 1996). The 2OG and Fe(II)-dependent dioxygenases (2OGD) are the second largest family of plant enzymes and are also widespread in fungi, mammals, and microorganisms. 2OGDs also play roles in various oxygenation/hydroxylation reactions (Kawai et al., 2014). The 2OGD superfamily has been shown to participate in antibiotic biosynthesis, collagen biosynthesis, DNA repair, and stress responses (Aravind and Koonin, 2001).

The 2OGOs play a wide variety of biological roles in bacteria, plants, and human (Welford et al., 2005). 2OGOs function in adventitious root formation of apple trees (Butler and Gallagher, 1999), mugineic acid synthesis in barley (Okumura et al., 1994), and fruit ripening in tomato (Trentmann and Kende, 1995). 2OGs have been reported to regulate metabolic activities such as lipid absorption/metabolism, muscle performance, and cancerogenesis in the human body (Harrison and Pierzynowski, 2008), as well as nitrogen metabolism in *Rhodospirillum rubrum* (Teixeira et al., 2010). Cheng et al. (2014) also reported the function of four 2OG dependent oxygenases, anthocyanidin synthase (ANS), flavonol synthase (FLS), flavone synthase I (FNS I), and flavanone 3- β -hydroxylase (FHT), in plant flavonoid biosynthesis, as well as the relationship between their catalytic activity and tertiary structure in Apiaceae species. Finally, Araujo et al. (2012) identified the effects of 2OG on 2OGD gene expression in *Arabidopsis* leaves.

In a previous study, 34 2-oxoglutarate oxygenase (2OGO) transcripts with different expression levels were obtained from transcriptome data of *V. labruscana* treated with low temperatures (Ahn et al., 2014). We also identified six 2OGO genes in *V. labruscana* by comparing structures, nucleotide sequences, and deduced amino acid sequences. In addition, we analyzed the differential expression patterns in response to exposure to low temperature for 1–4 weeks in ‘Campbell Early’ grapevines.

Materials and Methods

Plant Materials and Cold Treatment

Grapevines of ‘Campbell Early’ with green shoots grown in a greenhouse were maintained at 4°C in a growth chamber under a 16:8 photoperiod for 0, 1, 2, 3, and 4 weeks. Healthy green leaves of grapevines were harvested at the indicated times, immediately frozen in liquid nitrogen and stored at -80°C for further analysis.

RNA Isolation and Real-time PCR Analysis

Total RNA was extracted from grapevine leaves harvested at each time point using the method described by Chang et al. (1993). First-strand cDNA was synthesized from the total RNA (500 ng) using a GoScript Reverse Transcriptome System (Promega, Madison, WI, USA). Real time PCR was conducted with a C1000TM Thermal Cycler (BioRad, Hercules, CA, USA) using SYBR Premix Ex (Takara Bio Inc., Osaka, Japan) as the fluorescent dye, then analyzed using the CFX Manager software (BioRad). The PCR amplification was conducted by initial denaturation at 95°C for 30 s, followed by 40 cycles of 95°C for 5 s and 60°C for 30 s. Transcripts levels were normalized using the standard-curve method and calculated against

Table 1. Sequences and NABIC IDs of primers used for real-time PCR analysis.

Gene	NABIC ^z ID	Primer sequences
VICE2OGO1 ^y	NS-0795-000001	5'-CAACCTTGGGGATCAGATTCAG-3' 5'-AGTGAGACGCGCTCTTTTATGG-3'
VICE2OGO2	NS-0796-000001	5'-TGGGTTCATCGTTTCTATGGT-3' 5'-GAAGGAAGGAGCTCCGGTGTAT-3'
VICE2OGO3	NS-0797-000001	5'-GAGAGACGGAGCCAGTTCTCAG-3' 5'-GCCTGATTCGGAGTTTCTTTCA-3'
VICE2OGO4	NS-0798-000001	5'-CGTGGATCTCCTAGTCGCTGAT-3' 5'-ATTCTCCGAACGACTTCCAGTG-3'
VICE2OGO5	NS-0799-000001	5'-CTTCCATCTCTTTCTCCGACCA-3' 5'-AGTTGGAAGAAGCCGTGTTGAG-3'
VICE2OGO6	NS-0800-000001	5'-ATCCCTGCAAAGCTACGTCAAG-3' 5'-GCTGGAGTGCTTCTTGGTC-3'

^zNABIC–National Agricultural Biotechnology Information Center, RDA, Korea.

^yVICE2OGO : 2-Oxoglutarate (2OG) and Fe(II) oxygenase genes from *Vitis labruscana* cv ‘Campbell Early’.

the grapevine *Actin* gene (AB372563) as an internal control (Kim et al., 2014). The specific primer sequences of six *2OGO* genes were designed by alignment of nucleotide sequences using Primer 3 (<http://frodo.wi.mit.edu/primer3>) for real-time PCR (Table 1), and used to investigate their expression in response to low temperature in grapevines.

Cloning of 2OG Genes and Sequence Analysis

Among 34 DEGs identified from transcripts of ‘Campbell Early’ kept at 4°C, by next generation sequencing (NGS), six loci showing different transcription levels with more than 100 scores were selected to characterize their structures and to evaluate their expressions against chilling treatment. Nucleotide and amino acid sequence data of *VICE2OGO1-VICE2OGO6* (*Vitis labruscana* 2-oxoglutarate (2OG) and Fe(II) oxygenase 1-6) were deposited in the National Agricultural Biotechnology Information Center (NABIC), Rural Development Administration Korea database under accession numbers NS-0795-000001-NS-0800-000001 (Table 1), respectively.

Comparison of Gene Structure

Amino acid sequences were deduced from nucleotide sequences using the Translate program hosted by the ExPASy Proteomics Server (<http://web.expasy.org/translate/>). The primary structure of genes was performed using ProtParam (<http://web.expasy.org/protparam/>), while the secondary structure was analyzed using the Self-Optimized Prediction Method with Alignment (SOPMA) (https://npsa-prabi.ibcp.fr/cgi-bin/npsa_automat.pl?page=npsa_sopma.html). The protein domain was analyzed using the Simple Modular Architecture Research Tool program (SMART) (<http://smart.embl-heidelberg.de/>) (Letunic et al., 2014). Multiple alignment and pairwise comparisons of amino acid sequences were performed using T-coffee (<http://tcoffee.vital-it.ch/apps/tcoffee/do:regular>) (Notredame et al., 2000). A phylogenetic tree was constructed by the neighbor-joining method using MEGA version 6.0 (The Biodesign Institute, Tempe, USA).

Results and Discussion

Among six transcripts from ‘Campbell Early’ grapevine (*V. labruscana*), the highest similarity (38.6%) was observed between *VICE2OGO1* and *VICE2OGO2*, while the lowest similarity (19.2%) was between *VICE2OGO2* and *VICE2OGO3* based on deduced amino acid sequences (Table 2). The deduced amino acid sequences of *VICE2OGO2*, *VICE2OGO1*, *VICE2OGO6*, and *VICE2OGO4* showed 34.0, 33.7, 30.1, and 29.3% sequence similarity with those of *VICE2OGO4*, *VICE2OGO4*, *VICE2OGO2*, and *VICE2OGO6*, respectively.

The primary structures of the six *VICE2OGO* genes of ‘Campbell Early’ were characterized (Table 3). The six *VICE2OGO* transcripts ranged from 2298 to 1053 bp in size. The sizes of the *2OGO* genes varied in position and the sizes of the open reading frame (ORF) ranged from 92 to 1177, 57 to 1103, 1 to 1052, 109 to 1152, 47 to 1188, and 52 to 1002 nucleotides, respectively. The results showed that the size of the six *2OGO* proteins varied from 316 to 380 amino acids (35.80 to 41.02 kDa). *VICE2OGO4* was identified as stable (II < 40) based on the instability index (II) proposed by Guruprasad et al. (1990).

The aliphatic index (AI) of a protein is a measure of the relative content occupied by the aliphatic residue side chain (alanine, valine, leucine, and isoleucine) (Gasteiger et al., 2005). AI index of the *VICE2OGOs* varied from 78.32 to 91.61, which is regarded as a positive factor for the increase in thermal stability of globular proteins in grapevines. The grand average hydropathy (GRAVY) value for a protein is calculated as the sum of hydropathy values of all the amino acids divided by the number of residues in the sequence (Kyte and Doolittle, 1982). The GRAVY index of *VICE2OGO* proteins ranges from -0.434 to -2.210, indicating that they are hydrophilic throughout the length of the protein.

Secondary structure prediction of these proteins was conducted using SOPMA (Table 4). The results showed that alpha helixes, extended strands, beta turns, and random coils varied from 23.95 to 41.71%, 16.00 to 22.34%, 6.65 to 9.22%, and

Table 2. Identity of amino acid sequences of 2-oxoglutarate oxygenase (*2OGO*) proteins of *Vitis labruscana*.

	VICE2OG1 ²	VICE2OG2	VICE2OG3	VICE2OG4	VICE2OG5	VICE2OG6
VICE2OGO1		38.6	29.0	33.7	27.2	25.7
VICE2OGO2	38.6		25.0	34.0	22.0	30.1
VICE2OGO3	20.9	19.2		19.9	26.2	23.3
VICE2OGO4	33.7	34.0	19.9		21.7	29.3
VICE2OGO5	25.4	21.8	26.2	22.6		25.9
VICE2OGO6	25.7	30.1	23.3	29.3	25.9	

²VICE2OGO: *2OGO* proteins of *Vitis labruscana* cv ‘Campbell Early’.

Table 3. Primary structure analysis of 2-oxoglutarate oxygenase (*2OGO*) proteins of *Vitis labruscana*.

Gene	Nucleotide length (bp)	ORF ² (bp)	Protein length (aa)	Molecular mass (kDa)	Isoelectric point (pI)	Instability index (II)	Aliphatic index (AI)	Grand Average Hydropathy
VICE2OGO1	1376	1086	367	41.47	6.93	41.43	91.61	-0.275
VICE2OGO2	2298	1047	348	39.57	5.79	40.10	82.82	-0.409
VICE2OGO3	1053	1053	350	39.50	5.81	57.44	89.43	-0.288
VICE2OGO4	1340	1044	347	39.73	5.47	37.71	87.06	-0.434
VICE2OGO5	1189	1143	380	41.02	5.77	47.08	78.32	-0.341
VICE2OGO6	1331	951	316	35.80	4.99	43.81	85.44	-0.210

²ORF: open reading frame.

Table 4. Secondary structure prediction of 2-oxoglutarate oxygenase (2OGO) proteins of *Vitis labruscana* by SOPMA.

Protein	Alpha helix (%)	Extended strand (%)	Beta turn (%)	Random coil (%)
VICE2OGO1	35.97	22.34	8.72	32.97
VICE2OGO2	33.05	18.39	8.33	40.23
VICE2OGO3	41.71	16.00	7.14	35.14
VICE2OGO4	35.45	20.17	9.22	35.16
VICE2OGO5	23.95	16.84	7.63	51.58
VICE2OGO6	40.51	18.99	6.65	33.86

32.97 to 51.58 %, respectively. Analysis of the structure of predicted proteins showed that most proteins have Pfam domains, including non-haem dioxygenase in the morphine synthesis N-terminal (DIOX_N) and 2OG-Fe(II) oxygenase superfamily (2OG-Fe-Oxy), which are conserved and essential domains of 2OGOs (Fig. 1).

Transmembrane helix (TH) was only detected in VICE2OGO3. VICE2OGO3 and VICE2OGO5 contained the structural classification of proteins (SCOP) domain. Multiple alignments of the amino acid sequences deduced from the nucleotide sequences of six 2OGO genes in the ‘Campbell Early’ grapevine are shown in Fig. 2.

A phylogenetic tree based on the amino acid sequences of 2OGO transcripts was constructed to investigate the genetic relationships of VICE2OGOs with other plant proteins (Fig. 3). The 2OGO transcripts were divided into two groups, one including *VICE2OG3* and another group comprising the remaining transcripts. Among *VICE2OGOs*, the sequence of *VICE2OGO2* showed a high level of similarity to *At2OGO* (NM_117118.4).

Expression of genes was determined by real-time PCR of grapevine leaves treated with low temperature using specific

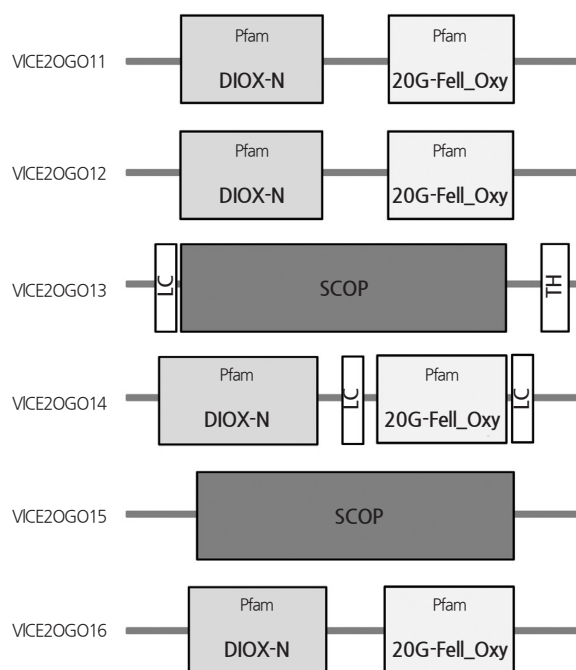


Fig. 1. Structural analysis of the 2-oxoglutarate oxygenase (2OGO) proteins of *Vitis labruscana*. DIOX_N, non-haem dioxygenase in morphine synthesis N-terminal; 2OG-Fell_Oxy, 2OG-Fe(II) oxygenase superfamily; LC, low complexity; SCOP, structural classification of proteins; TH, transmembrane helix.

VICE2OG1	70	DARLRQATLDRI	SGACREWGFQV	VNHGVRPELMK	SIRQLWRDFFHL	PLEAKQV-YAN	SPATYEGYGS	RGLGVEKGAK	LWDSYF	152
VICE2OG2	56	---GRP	DVVKIQACQ	HDGFFQIK	NHGVSETMIS	NMLRLARDF	FLPESE	RKNYS	DNPSKTTRL	136
VICE2OG3	78	---RD	GDSIRRI	LSARACG	VFRINELG	VADQLQSI	VAAE	ESVFRV	SEEVKTK-F	144
VICE2OG4	38	---AV	DLLVADIG	KACEEWG	FFKVINH	GVPLEV	VRRIEA	AMKEFF	DQSID	115
VICE2OG5	71	---Q	NLNLLS	DLFS	SQHGFF	QLIDHP	IPSQL	ARS	AESESL	145
VICE2OG6	28	---E	WKGLR	SKVRE	ACETY	GCFGL	VCD	AI	PAKLR	101
VICE2OG1	153	FLNY-----	L--PE---	SAR-DEN	KWPTT	PSCREL	VHEYS	KAVVEL	CGILMK	216
VICE2OG2	137	RLHC-----	Y--P---	LED-YV	HEWPS	NPPTF	FRED	VAEY	CTSTRE	197
VICE2OG3	145	WLR-----	-----	EE-EM	VEWAE	EAI-G	CEI	YQS	FSKM	208
VICE2OG4	116	DI	AVED	PLVIS	ASP	DPDQ	ELTK-	Y	YNQW	190
VICE2OG5	146	CLES-----	-----	SC	STEST	HL	SLSL	REF	T	198
VICE2OG6	102	GIDD-----	A--P	QPQA	QAFT	NLLW	PEGN	PAFC	GALIS	166
VICE2OG1	217	GACLR	VNY	PKCP	QPD	LT	LGIS	PHSDP	--G	295
VICE2OG2	198	GQH	MAM	NY	PP	CP	PE	LT	YGLP	275
VICE2OG3	209	LSL	YR	NH	D	S	L	M	-D	287
VICE2OG4	191	TS	F	RL	NH	Y	PP	CP	W	270
VICE2OG5	199	C	P	L	W	I	S	E	D	272
VICE2OG6	167	AS	V	R	L	M	K	Y	T	245
VICE2OG1	296	EHR	V	V	N	S	I	K	E	358
VICE2OG2	276	LH	R	A	V	N	C	N	K	338
VICE2OG3	288	S	G	E	F	I	F	D	S	327
VICE2OG4	271	E	H	R	V	V	A	N	S	332
VICE2OG5	273	R	G	R	A	V	N	R	G	355
VICE2OG6	246	R	H	R	V	M	N	G	D	303

Fig. 2. Multiple alignment of amino acid sequences of unique regions from 2-oxoglutarate oxygenases (2OGOs). VICE2OGO: 2OGO proteins of *Vitis labruscana* cv. 'Campbell Early'.

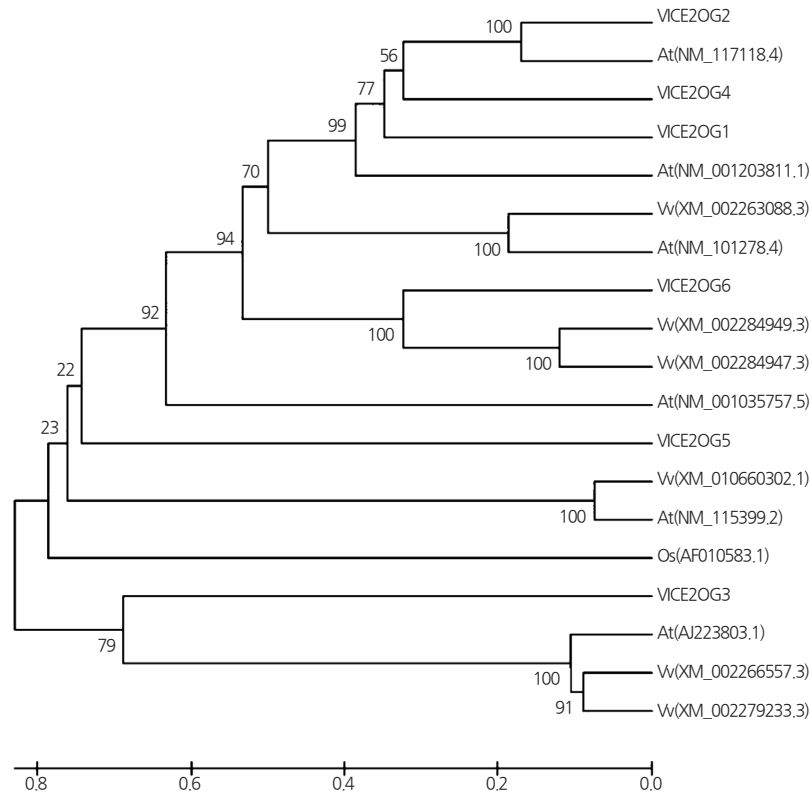


Fig. 3. Phylogenetic tree of 2-oxoglutarate oxygenase (VICE2OGO) proteins from *Vitis labruscana* transcripts with 2OGO proteins from *Vitis vinifera* (Vv), *Arabidopsis* (At), and rice (Os).

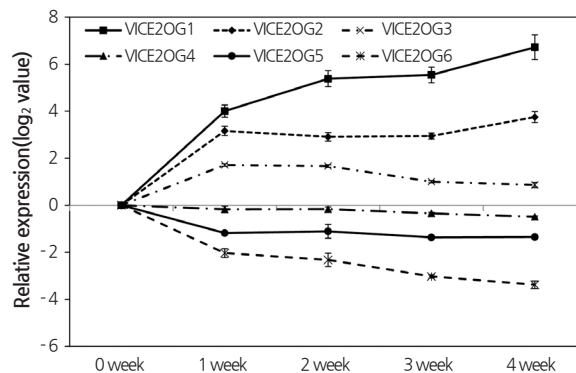


Fig. 4. Expression of 2-oxoglutarate oxygenase (*2OGO*) genes by quantitative real-time PCR analysis of ‘Campbell Early’ grapevine leaves treated with cold. Vertical bars indicate the standard deviation of three replicates. *VICE2OGO*: *2OGO* genes from *Vitis labruscana* cv. ‘Campbell Early’.

primers for each *VICE2OGO* gene (Fig. 4). *VICE2OGO1*, *VICE2OGO2*, and *VICE2OGO3* genes were up-regulated, whereas *VICE2OGO4*, *VICE2OGO5*, and *VICE2OGO6* genes were down-regulated compared to controls in grapevines subjected to low temperature. Differential expression patterns from the real-time PCR analysis were consistent with those observed upon transcript analysis of grapevines maintained at low temperature for 4 weeks.

Low temperature is a serious abiotic stress that causes physiological damage and reduced productivity in growing grapevines. In the present study, we identified structural characteristics and analyzed the expression of six *2OGOs* of ‘Campbell Early’ grapevine leaves in response to low temperature. The differential expression pattern of six *2OGOs* was detected by real time PCR analysis with gene specific primers in ‘Campbell Early’ grapevines exposed to chilling temperature. Kim et al. (2014) reported that six basic helix-loop-helix (bHLH) genes showed up- and down-regulated expression patterns in response to low temperature treatments in ‘Campbell Early’ and ‘Muscat Bailey A’ grapevine leaves and latent buds.

2OG is related to the biosynthesis and secondary metabolism of signaling molecules such as alkaloids, gibberellins, and biosynthesis of flavonoids. Kawai et al. (2014) reported that *2OG* Fe(II) dependent oxygenases were extensively present in various plant species. In *Arabidopsis*, more than 100 members of *2OGOs* have been identified, which is greater than the number of identified and classified *2OGOs* based on sequence analysis in a previous report (Presscot and Lloyd, 2000). Phylogenetic analysis showed that the six *VICE2OGO* and other *2OGO* genes were distributed in two major groups within several clades in the present study.

It has been reported that *2OGO* was down-regulated throughout the cold treatment period and encoded a protein that demonstrated 44% identity with that of rice (Ciannamea et al., 2006). A hydropathy plot revealed that the predicted OsLti (*Oryza sativa* low-temperature-induced gene) proteins, OsLti6a and OsLti6b, were highly hydrophobic throughout the length of the protein, with GRAVY values of 1.51 and 1.42, respectively (Morsy et al., 2005).

Detection of *2OGD* sequences has been reported in various plants by searching for the *2OG*-Fe(II)-Oxy motif against whole amino acid sequences, with 142, 130, 114, and 74 *2OGDs* reported in the *Picea*, *Arabidopsis*, *Oryza*, and *Selaginella* genomes, respectively (Kawai et al., 2014). The presence of typical domain positions at 38–499 of the *2OG*-Fe(II) oxygenase superfamily was detected in tea plants by ExPASy, BLAST, and SMART analyses (Rani et al., 2012). The flexibility and stability of *2OGO* proteins have been influenced by dehydration and protein structure at random coils in response to water

stress (Buxbaum, 2007; Qu et al., 1998). Schofield and Zhang (1999) suggested that biasing the reaction pathway of 2OG-dependent oxygenases to deliberately generate radicals might lead to inhibition. A 2-OGD superfamily protein containing a 2OG-Fe(II)-oxygenase domain shared approximately 67% amino acid identity with downy mildew resistant 6 (*DMR6*) in *Arabidopsis*, and expression of *DMR6* induced the expression of a subset of defense-associated genes such as pathogenesis-related genes against foreign stress (Damme et al., 2008). 2OGs were shown to have an activity of flavanone 3beta-hydroxylase (F3H) in ethylene synthesis and the biosynthesis of flavanoids, catechins and anthocyanidins associated with defense responses to stress in plants including wheat (Giovanini et al., 2006) and petunia (Lukacin and Britsch, 1997).

In this study, we analyzed the structure, nucleotide sequences and deduced amino acid sequences of six *2OGO* genes selected by transcriptome analysis by NGS of ‘Campbell Early’ grapevine leaves exposed to low temperature. All tested *VICE2OGO* genes showed differential expression in response to low temperature treatment in ‘Campbell Early’ grapevine leaves. It suggests that induction of *VICE2OGO* genes in expression level were related with defense responses to low temperature in grapevines. Further studies on gene expression based on transcriptome analysis in various organs like root and could provide meaningful information regarding the response of grapevine to stress from low temperatures in the future. The information presented herein will contribute to elucidation of mechanisms of cold hardness and selection of valuable molecular genetic resources responding to low temperature in grapevines.

Literature Cited

- Ahn, S.Y., S.A. Kim, S.H. Jo, and H.K. Yun. 2014. De novo transcriptome assembly of *Vitis flexuosa* grapevines inoculated with *Elsinoe ampelina*. *Plant Genet. Res.* 12:5130-5133.
- Araujo, W.L., T. Tohge, A. Nunes-Nesi, D.M. Daloso, M. Nimick, I. Krahnert, V.I. Bunik, G.B. Moorhead, and A.R. Fernie. 2012. Phosphonate analogs of 2-oxoglutarate perturb metabolism and gene expression in illuminated *Arabidopsis* leaves. *Front Plant Sci* 3:114.
- Aravind, L. and E.V. Koonin. 2001. The DNA-repair protein AlkB, EGL-9, and leprecan define new families of 2-oxoglutarate- and iron-dependent dioxygenases. *Genome Biol.* 2:1-0007.
- Butler, E.D. and T.F. Gallagher. 2000. Characterization of auxin-induced ARRO-1 expression in the primary root of *Malus domestica*. *J. Exp. Bot.* 51:1765-1766.
- Buxbaum, E. 2007. Protein structure. *Fundamentals of protein structure and function*, p. 12-37. Springer Science Business Media, New York, USA.
- Chang, S., J. Puryear, and J. Cairney. 1993. A simple and efficient method for isolating RNA from pine trees. *Plant Mol. Biol.* 11:113-116.
- Cheng, A.X., X.J. Han, Y.F. Wu, and H.X. Lou. 2014. The function and catalysis of 2-oxoglutarate-dependent oxygenases involved in plant flavonoid biosynthesis. *Int. J. Mol. Sci.* 15:1080-1095.
- Ciannamea, S., J. Busscher-Lange, S. de Folter, G.C. Angent, and R.G.H. Immink. 2006. Characterization of the vernalization response in *Lolium perenne* by a cDNA microarray approach. *Plant Cell Physiol.* 47:481-492.
- Damme, M., R.P. Huibers, J. Elberse, and G.V. Ackerveken. 2008. *Arabidopsis* *DMR6* encodes a putative 2OG-Fe(II) oxygenase that is defense-associated but required for susceptibility to *downy mildew*. *Plant J.* 54:785-793.
- Gasteiger, E., C. Hoogland, A. Gattiker, S. Duvaud, M.R. Wilkins, R.D. Appel, and A. Bairoch. 2005. Protein identification and analysis tools on the ExPASy server, p. 571-607. In: J.M. Walker (ed.). *The proteomics protocols handbook*. Humana press, Totowa, NJ.
- Giovanini, M.P., D.P. Puthoff, J.A. Nemacheck, O. Mittapalli, K.D. Saltzmann, H.W. Ohm, R.H. Shukle, and C.E. Williams. 2006. Gene-for-gene defense of wheat against the Hessian fly lacks a classical oxidative burst. *Mol. Plant Microbe Interact.* 19:1023-1033.
- Guruprasad, K., B.V. Reddy, and M.W. Pandit. 1990. Correlation between stability of a protein and its dipeptide composition: a novel approach for predicting in vivo stability of a protein from its primary sequence. *Protein Eng.* 4:155-161.
- Harrison, A.P. and S.G. Pierzynowski. 2008. Biological effects of 2-oxoglutarate with particular emphasis on the regulation of protein, mineral and lipid absorption/metabolism, muscle performance, kidney function, bone formation and cancerogenesis, all viewed from a healthy ageing perspective state of the art. *J. Physiol. Pharmacol.* 59(Suppl. 1):91-106.
- Kawai, Y., E. Ono, and M. Mizutani. 2014. Evolution and diversity of the 2-oxoglutarate-dependent dioxygenase superfamily in plants. *Plant J.* 78:328-343.
- Kim, S.A., S.Y. Ahn, S.H. Kim, J.H. Han, and H.K. Yun. 2014. Expression of basic helix-loop-helix transcripts during low temperature treatment in grapevines. *Plant Breed. Biotechnol.* 2:110-116.

- Kyte, J. and R.F. Doolittle. 1982. A simple method for displaying the hydropathic character of a protein. *J. Mol. Biol.* 157:105-132.
- Letunic, I., T. Doerks, and P. Bork. 2014. SMART: recent updates, new developments and status in 2015. *Nucl. Acids Res.* 43:257-260.
- Lukacin, R. and L. Britsch. 1997. Identification of strictly conserved histidine and arginine residues as part of the active site in *Petunia hybrida* flavanone 3 β -hydroxylase. *Eur. J. Biochem.* 249:748-757.
- Morsy, M.R., A.M. Almutairi, J. Gibbons, S.J. Yun, and G. Benildo. 2005. The *OsLti6* genes encoding low-molecular-weight membrane proteins are differentially expressed in rice cultivars with contrasting sensitivity to low temperature. *Gene* 344:171-180.
- Nam, J.C., S.J. Park, S.M. Jeong, J.H. Noh, Y.Y. Hur, and K.S. Park. 2012. Threshold of winter injury according to low temperature in dormancy and growing stage in several grape (*Vitis* hybrid) cultivars. *Korean J. Hortic. Sci. Technol.* 30(Suppl. II):100-101. (Abstr.)
- Notredame, C., D.G. Higgins, and J. Heringa. 2000. T-Coffee: A novel method for fast and accurate multiple sequence alignment. *J. Mol. Biol.* 302:205-217.
- Okumura, N., N.K. Nishizawa, Y. Umehara, T. Ohata, H. Nakanishi, T. Yamaguchi, M. Chino, and S. Moil. 1994. A dioxygenase gene (*lds2*) expressed under iron deficiency conditions in the roots of *Hordeum vulgare*. *Plant Mol. Biol.* 25:705-719.
- Park, G.H., J.W. Lim, and G.J. Lee. 2000. Management of chilling injury in grapevine shoots in unheated plastic house in Korea. 2000 Research Report, p. 388-397. Gyeonggi-do Agricultural Research and Extension Services, Korea.
- Prescott, A.G. and M.D. Lloyd. 2000. The iron(II) and 2-oxoacid-dependent dioxygenases and their role in metabolism. *Nat. Prod. Rep.* 17:367-383.
- Prescott, A.G. and P. John. 1996. Dioxygenases: molecular structure and role in plant metabolism. *Annu. Rev. Plant Physiol. Plant Mol. Biol.* 47:245-271.
- Qu, Y., C.L. Bolen, and D.W. Bolen. 1998. Osmolyte-driven contraction of a random coil protein. *Proc. Natl. Acad. Sci. USA.* 95:9268-9273.
- Rani, A., K. Singh, P.S. Ahuja, and S. Kumar. 2012. Molecular regulation of catechins biosynthesis in tea [*Camellia sinensis* (L.) O. Kuntze]. *Gene* 495:205-210.
- Schofield, S.J. and Z. Zhang. 1999. Structural and mechanistic studies on 2-oxoglutarate-dependent oxygenases and related enzymes. *Curr. Opin. Struct. Biol.* 9:722-731.
- Tattersall, E.A., J. Grimplet, L. DeLuc, M.D. Wheatley, D. Vincent, C. Osborne, A. Ergul, E. Lomen, R.R. Blank, K.A. Schlauch, J.C. Cushman, and G.R. Cramer. 2007. Transcript abundance profiles reveal larger and more complex responses of grapevine to chilling compared to osmotic and salinity stress. *Funct. Integr. Genomics* 7:317-333.
- Teixeira, P.F., T.T. Selao, V. Henriksson, H. Wang, A. Noren, and S. Nordlund. 2010. Diazotrophic growth of *Rhodospirillum rubrum* with 2-oxoglutarate as sole carbon source affects regulation of nitrogen metabolism as well as the soluble proteome. *Res. Microbiol.* 161:651-659.
- Trentmann, S.M. and H. Kende. 1995. Analysis of Arabidopsis cDNA that shows homology to the tomato E8 cDNA. *Plant Mol. Biol.* 29:161-166.
- Warmund, M.R., P. Guinan, and G. Fernandez. 2008. Temperatures and cold damage to small fruit crops across the Eastern United States associated with the April 2007 freeze. *HortScience* 43:1643-1647.
- Welford, R. W., J. M. Kirkpatrick, L.A. McNeill, M. Puri, N.J. Oldham, and C.J. Schofield. 2005. Incorporation of oxygen into the succinate co-product of iron (II) and 2-oxoglutarate dependent oxygenases from bacteria, plants and humans. *FEBS Lett.* 579:5170-5174.
- Zhao, M.X., Y.L. Jiang, Y.X. He, Y.F. Chen, Y.B. Teng, Y. Chen, C.C. Zhang, and C.Z. Zhou. 2010. Structural basis for the allosteric control of the global transcription factor NtcA by the nitrogen starvation signal 2-oxoglutarate. *Proc. Natl. Acad. Sci. USA.* 107:12487-12492.